



## 生物实验室

## 基于非数据依赖的鞘脂菌蛋白质组学分析方法的建立

窦玥<sup>1</sup> 熊为亮<sup>2</sup> 梁如冰<sup>2</sup> 侯敬丽<sup>\*3</sup>

1 上海交通大学药学院 上海 200240

2 上海交大学生命科学技术学院 上海 200240

3 上海交通大学分析测试中心 上海 200240

**摘要:**【背景】鞘脂菌是一类可高效降解以菲为代表的多环芳烃有机污染物的菌株,其在环境污染治理及生物技术领域具有广阔的应用前景。【目的】为了优化测试方法,获得更完整的鞘脂菌 *Sphingobium yanoikuyae* SJTF-8 在菲胁迫下表达差异的蛋白。【方法】利用数据依赖型及数据非依赖型两种蛋白质组学数据采集方法,比较了鞘脂菌 SJTF-8 在菲胁迫下蛋白质水平的表达变化。【结果】两种技术方法下共得到 580 个表达差异蛋白,这些蛋白在细胞代谢、转运和调控等方面发挥一定功能。【结论】数据非依赖性采集(data-independent acquisition, DIA)技术在重复性以及低丰度蛋白的检测上明显好于数据依赖型采集(data dependent acquisition, DDA)技术,因此, DIA 在实际可用的表达差异蛋白检出方面具备明显优势,为发现菲胁迫下细胞诱导表达的低丰度调控蛋白提供帮助。

**关键词:** 数据非依赖性质谱采集方法, 数据依赖性质谱采集方法, 菲, 鞘脂菌

## Establishment of a data-independent acquisition proteomic analysis method for *Sphingobium yanoikuyae*

DOU Yue<sup>1</sup> XIONG Wei-Liang<sup>2</sup> LIANG Ru-Bing<sup>2</sup> HOU Jing-Li<sup>\*3</sup>

1 School of Pharmacy, Shanghai Jiao Tong University, Shanghai 200240, China

2 School of Life Science and Biotechnology, Shanghai Jiao Tong University, Shanghai 200240, China

3 Instrumental Analysis Center of Shanghai Jiao Tong University, Shanghai 200240, China

**Abstract:** [Background] *Sphingobium* is a kind of bacteria which can effectively degrade polycyclic aromatic hydrocarbons (PAHs) represented by phenanthrene. This characteristic makes *sphingobium* have broad application prospects in the field of environmental pollution control and biotechnology. [Objective] The aim of this study is to optimize the quantification method in order to acquire the complete differentially expressed proteins of *Sphingobium yanoikuyae* SJTF8 under phenanthrene stress. [Methods] Data dependent acquisition (DDA) and data-independent acquisition (DIA) proteomics analysis methods were used in this experiment. [Results] These two methods had commonly quantified 580 differentially expressed proteins, which played important roles in cell metabolism, transport and regulation. [Conclusion] DIA has a significant advantage in the quantification of differentially expressed proteins, which is helpful for the discovery of low abundance regulatory proteins induced by phenanthrene stress.

**Keywords:** Data-independent acquisition, Data dependent acquisition, Phenanthrene, *Sphingobium yanoikuyae*

\*Corresponding author: E-mail: hopie00@sjtu.edu.cn

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\*通信作者: E-mail: hopie00@sjtu.edu.cn

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多环芳烃(PAHs)是一种普遍存在于生态系统中的全球性污染物, 由于其具有潜在的毒性、致癌性、致畸性和致突变性, 因而受到人们的广泛关注<sup>[1]</sup>。微生物法修复环境中多环芳烃污染是一种被广泛研究的手段<sup>[2]</sup>。菲作为美国环境保护署重点关注的污染物之一, 已被广泛用作研究这些微生物降解多环芳烃的模型物质<sup>[3]</sup>。菲是一种低分子量、具有 3 个苯环结构的疏水性多环芳烃。菲的化学性质非常稳定, 易在环境中沉积, 并通过食物链向人体或者动物富集使细胞产生癌变<sup>[4]</sup>。

数据非依赖性采集(data-independent acquisition, DIA)技术是近几年新出现的蛋白质组学非标记定量技术, 最早由 Venable 在 2004 年提出<sup>[5]</sup>。不同于传统的需要选择特定母离子进行碎裂的数据依赖型采集(data dependent acquisition, DDA)技术, DIA 可采集所有的碎片离子信息, 从而具有全景式扫描、数据可回溯等优势<sup>[6]</sup>。得益于数据分析和质谱采集速度、质量精度和分辨率的显著进步, 近几年来 DIA 技术发展迅速, 涌现出大量新的 DIA 优化改进技术和多个自动化 DIA 数据分析工具<sup>[7]</sup>。高质量的 DIA 定量技术作为蛋白质组定量新技术将会被逐渐推广并应用于生命科学和医学等不同领域的研究中<sup>[8-9]</sup>。

本实验室长期从事微生物环境修复菌的筛选和降解调控机制的研究。我们在土壤中筛选得到一株可高效降解菲的鞘脂菌(*Sphingobium yanoikuyae* strain SJTF-8), 为了进一步比较 DDA 和 DIA 定量技术的特点和在蛋白质组学中的应用范围, 本研究以这株可高效降解菲的鞘脂菌为研究对象, 利用 DDA 和 DIA 两种蛋白质组学定量技术比较鞘脂菌 SJTF-8 在菲胁迫下蛋白质水平的表达变化, 以期发现更多参与多环芳烃降解和调控的蛋白, 为微生物降解多环芳烃机制的研究提供理论依据。

## 1 材料与方法

### 1.1 样品

实验菌株鞘脂菌 SJTF-8 由上海交通大学梁如冰实验室分离得到, 在菲中生长良好。

### 1.2 培养基

基础盐培养基(g/L):  $\text{KH}_2\text{PO}_4$  4.500,  $\text{K}_2\text{HPO}_4$  13.750,  $(\text{NH}_4)_2\text{SO}_4$  2.000,  $\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$  0.160,  $\text{FeSO}_4 \cdot 7\text{H}_2\text{O}$  0.005,  $\text{CaCl}_2$  0.011,  $\text{MnCl}_2 \cdot 4\text{H}_2\text{O}$  0.002。

### 1.3 主要试剂和仪器

菲, Sigma 公司; 测序用胰酶, Promega 公司。非接触式超声破碎仪, Scientz 公司; XBridge Peptide BEH  $\text{C}_{18}$  色谱柱, Waters 公司; Ultimate3000 液相色谱仪、EASY-nLC 1200 纳升液相色谱仪、Q-Exactive Plus 质谱仪、离心浓缩仪, Thermo Fisher 公司。

### 1.4 实验方法

实验方法设计流程图详见图 1。

#### 1.4.1 细菌培养

采用 LB 培养基和含 250  $\mu\text{g/mL}$  菲的基础盐培养基培养细胞。鞘脂菌 SJTF-8 在 30  $^{\circ}\text{C}$ 、220 r/min 振荡培养, 12 000 r/min 离心 20 min 收集 LB 培养的鞘脂菌 SJTF-8, 用 1 $\times$ PBS 缓冲液洗涤 3 次, 将部分预培养细菌接种到含有菲的培养基中, 并按上述方法培养, 每种条件下培养 3 份样品。

#### 1.4.2 菌全蛋白的提取及方法优化

12 000 r/min 离心 20 min 收集培养细胞, 并用 1 $\times$ PBS 缓冲液洗涤 3 次。加入 600  $\mu\text{L}$  细胞裂解液(50 mmol/L  $\text{NH}_4\text{HCO}_3$ , 8 mol/L 尿素)充分悬浮细胞, 置于非接触式超声破碎仪中超声破碎。超声程序为: 效率 99%, 超声 10 s, 停止 5 s。为了测试非接触式超声破碎仪破碎鞘脂菌的效率, 优化鞘脂菌的提取方法, 细菌的超声时间分别设置为 15、30、45、60 min。不同时间超声获得的细胞裂解液分别于 4  $^{\circ}\text{C}$ 、12 000 r/min 离心 30 min 去除细胞碎片。用 BCA 法测定蛋白浓度。

#### 1.4.3 蛋白的酶解

在 37  $^{\circ}\text{C}$  下使用 1 mol/L DTT 将每个样品中的 50  $\mu\text{g}$  蛋白质还原 1 h。使用 1 mol/L 碘乙酰胺(IAA)在避光处室温下处理半胱氨酸残基 40 min。参考 FASP 方法, 使用测序级胰蛋白酶(Promega, USA)酶解蛋白过夜, 胰蛋白酶与蛋白质的质量比为 1:50<sup>[10]</sup>。12 000 r/min 离心 20 min 收集酶解后产生的多肽混合物, 用离心浓缩仪干燥样品, 溶解在 0.1% 甲酸溶液中待测。

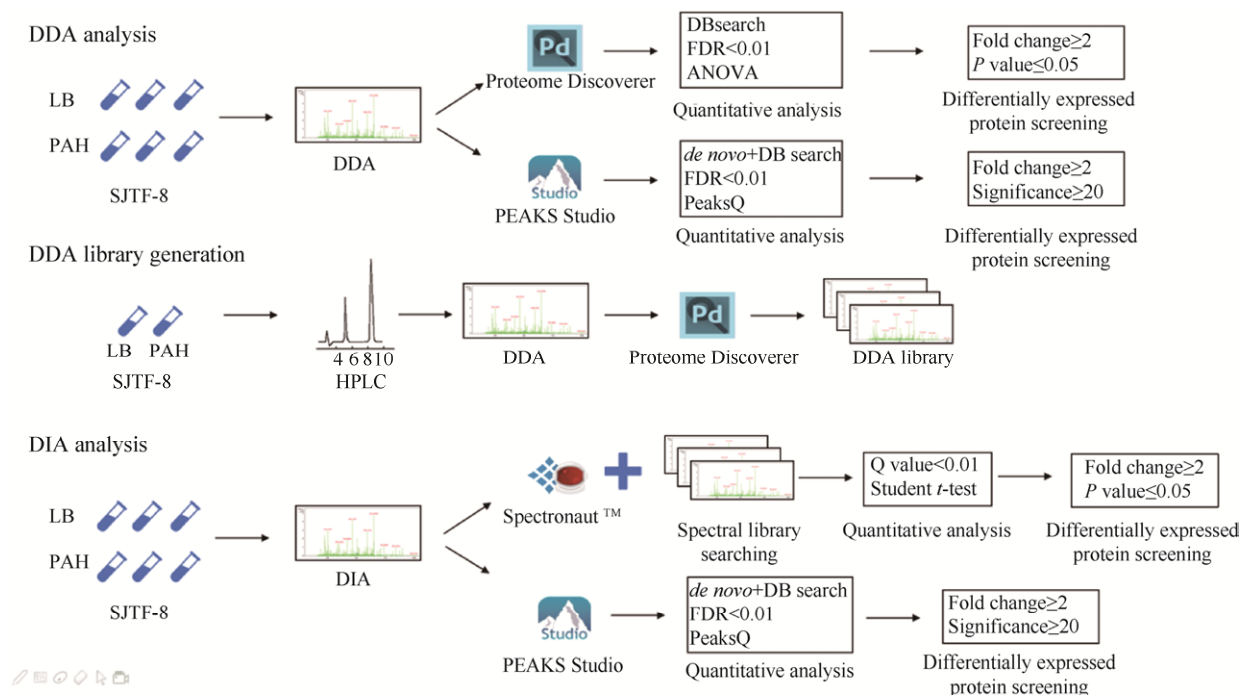


图1 质谱数据处理流程图

Figure 1 Mass spectrum data processing flow

#### 1.4.4 DDA 分析

酶解后的蛋白通过 Easy-nLC 1200 纳升液相色谱仪和含有纳米喷雾源的 Q-Exactive Plus 质谱仪进行分析。每个多肽样品用 0.1% 甲酸重新溶解,并在 C<sub>18</sub> 色谱柱(50 mm×15 cm, 1.7 μm)中分离。采用溶剂体系(流动相 A: 99.9% 水和 0.1% 甲酸; 流动相 B: 80% 乙腈和 0.1% 甲酸)分离多肽。以 200 nL/min 的流速使用 120 min 梯度洗脱肽,多肽洗脱梯度为: 0–2 min, 2%–6% B; 2–97 min, 6%–22% B; 97–107 min, 22%–32% B; 107–109 min, 32%–95% B; 109–119 min, 95% B; 119–120 min, 95%–2% B。质谱仪在数据依赖(DDA)模式下操作,在分辨率为 70 000 (AGC 3e6)的轨道阱中进行全扫描采集( $m/z$  350–1 500)。将分离出的前 20 个肽信号(电荷态≥+2)母离子通过高能碰撞(HCD)破碎,标准化碰撞能(normalized collision energy, NCE)为 28.0。毛细管的温度是 275 °C,喷雾电压是 1 900 V。子离子在分辨率为 17 500 (AGC 1e5)的轨道上测量。全扫描和 MS-MS 扫描的最大填充时间分别设置为 50 ms 和 45 ms,

动态排除时间设置为 30 s。

#### 1.4.5 DDA 谱图库的建立

取 200 μg 鞘脂菌蛋白的酶解产物通过高 pH 反相柱分离成 8 个组分。采用 XBridge Peptide BEH C18 色谱柱(250×4.6 mm, 5 μm, Waters),流动相 A 为 5 mmol/L 甲酸铵(pH 10.0),流动相 B 为 5 mmol/L 甲酸铵和 95% 乙腈混合液(pH 10.0),流速 0.8 mL/min,进样量 25 μL,柱温 40 °C,分离梯度为 0–5 min, 2% B; 5–6 min, 2%–5% B; 6–8 min, 5%–8% B; 8–43 min, 8%–25% B; 43–54 min, 25–40% B; 54–54.5 min, 40%–95% B; 54.5–59.5 min, 95% B; 59.5–60 min, 95%–2% B; 60–65 min, 2% B。将收集的溶液进行干燥,并重新溶解在 0.1% 甲酸中,用 nano-LC-MS/MS 进行分析,分析仪器及分析方法同 1.4.4。

将质谱产生的原始数据用 Proteome Discovery (PD, 2.3 版本)进行分析,生成谱图库。消化酶设置为 Trypsin,最大漏切设置为 2。母离子和子离子质量偏差分别设置成  $1 \times 10^{-5}$  和 0.02 Da。固定修饰

为 Carbamidomethyl (C), 可变修饰为 Oxidation (O), Deamidated (NQ)。使用 Target/Decoy 验证方法, 多肽和蛋白的 FDR 均设置为 1%。用 Spectronaut 11.0 (Biognosys AG 2013, 后文及表格中简写为 SPE) 打开 PD 生成的 .pdresult 搜库结果文件, 除不选择 iRT 校正外, 谱图库参数均按照软件默认设置。

1.4.6 DIA 分析

酶解后的多肽样品色谱分析方法同 1.4.4 DDA 分析。质谱仪在数据非依赖(DIA)模式下操作, 在分辨率为 70 000 (AGC 3e6)的轨道阱中进行全扫描采集( $m/z$  350–1 405), 见表 1 设置 30 个可变窗口, 全扫描 IT 为 50 ms。子离子在分辨率为 17 500 (AGC 1e6, IT 自动化, 默认带电量为 2)的轨道上测量。

表 1 DIA 隔离窗口设置

Table 1 DIA isolation window settings

| 窗口序号<br>Window No. | 开始质核比<br>Start m/z | 宽度<br>Width |
|--------------------|--------------------|-------------|
| 1                  | 372.5              | 34.5        |
| 2                  | 407                | 25          |
| 3                  | 432                | 21          |
| 4                  | 453                | 18          |
| 5                  | 471                | 18          |
| 6                  | 489                | 18          |
| 7                  | 507                | 18          |
| 8                  | 525                | 18          |
| 9                  | 543                | 18          |
| 10                 | 561                | 18          |
| 11                 | 579                | 18          |
| 12                 | 597                | 18          |
| 13                 | 615                | 18          |
| 14                 | 633                | 18          |
| 15                 | 651                | 18          |
| 16                 | 669                | 18          |
| 17                 | 687                | 18          |
| 18                 | 705                | 18          |
| 19                 | 723                | 18          |
| 20                 | 741                | 18          |
| 21                 | 759                | 18          |
| 22                 | 777                | 18          |
| 23                 | 795                | 22          |
| 24                 | 817                | 25          |
| 25                 | 842                | 25          |
| 26                 | 867                | 25          |
| 27                 | 892                | 42          |
| 28                 | 934                | 140         |
| 29                 | 1 074              | 220         |
| 30                 | 1 294              | No data     |

1.4.7 数据处理

(1) PD 中 DDA 数据处理(PD-DDA)

将 DDA 质谱原始数据导入 PD 搜索工具, 在峰识别后对每个样品进行 Label-free 定量分析, 使用 Uniprot\_SJTF-8 蛋白数据库(5 343 条目)进行搜库处理。设置如下: 消化酶设置为 Trypsin, 最大漏切设置为 2。母离子和子离子质量偏差分别设置成  $1 \times 10^{-5}$  和 0.02 Da。固定修饰为 Carbamidomethyl (C), 可变修饰为 Oxidation (O), Deamidated (NQ)。多肽和蛋白定性的 FDR 设置为 1%, 蛋白质的定量基于前 3 个肽段母离子峰面积。以 LB 培养细胞中的蛋白质为对照组进行无标记分析, 将蛋白质表达的倍数变化 2 或 0.5 和  $P \leq 0.05$  作为生物学效应的阈值。

(2) PEAKS 中 DDA 与 DIA 数据处理 (PEAKS-DDA, PEAKS-DIA)

将采集到的质谱数据导入 PEAKS Studio10.0 (Bioinformatics Solutions Inc.)中, 使用 Uniprot\_SJTF-8 蛋白数据库(5 343 条目)进行数据处理。将数据采集模式分别设置为 DDA 模式和 DIA 模式, 其余参数设置相同: 母离子和二级谱图的质量偏差分别设置成  $1 \times 10^{-5}$  和 0.02 Da; 胰酶的最大漏切位点数为 2; 固定修饰为 CarbamiDomethyl (C), 可变修饰为 Oxidation (O)、Deamidated (NQ); 多肽和蛋白定性的 FDR 设置为 1%。以 LB 培养细胞中的蛋白质为对照组进行无标记分析, 将蛋白质表达变化倍数  $\geq 2$  或  $\leq 0.5$  和 Significance  $\geq 20$  作为生物学效应的阈值。

(3) Spectronaut 中 DIA 数据处理(SPE-DIA)

将经 DIA 分析后的质谱数据导入 Spectronaut 11.0 (Biognosys AG 2013)中, 使用 PD 生成的谱图库进行数据处理。分析参数使用软件默认设置, 所有蛋白和肽定性分析在  $Q$  value 为 0.01 的水平下进行(相当于 FDR 为 1%), 在 MS2 水平下通过峰面积进行定量分析, 肽丰度通过对其各自的 MS2 片段离子的峰面积求和计算, 蛋白质丰度是通过对其各自的肽丰度求和计算。以 LB 培养细胞中的蛋白质为对照组进行无标记分析, 将蛋白质表达变化倍

数 $\geq 2$ 或 $\leq 0.5$ 和 $P \leq 0.05$ 作为生物学效应的阈值。

### 1.4.8 生物信息学分析

得到差异蛋白后,使用 Uniprot (<https://www.uniprot.org/uploadlists/>)和 String (version 11.0 <http://string-db.org/>),分别对差异蛋白进行 GO 分析和 KEGG 信号通路分析<sup>[11]</sup>。

## 2 结果与分析

### 2.1 鞘脂菌 SJTF-8 全蛋白提取效率优化

实验结果发现在含菲培养基中生长的鞘脂菌(菲细胞)生长过程中会产生大量胶状物质,与 LB 条件下培养获得的细胞(LB 细胞)相比,在相同的超声破碎条件下,菲细胞的破碎效率低于 LB 细胞。据此推测可能是因为鞘脂菌会分泌一种名叫结冷胶的胞外物质,导致了菲细胞破碎效率的降低<sup>[12-13]</sup>。因此专门对菲细胞破碎效率做了优化,如图 2 所示,破碎时间为 45 min 及以上的样品蛋白浓度是 30 min 及以下的近 2 倍,说明破碎时间对鞘脂菌 SJTF-8 细胞中的蛋白提取效率有影响,更长的超声时间有助于提高细胞破碎效率,超声时间超过 45 min 时,破碎效率没有明显变化。因此,选择使用 45 min 的超声时间来裂解细胞。

### 2.2 DDA 与 DIA 数据采集模式结果比较

#### 2.2.1 定性定量结果比较

分别整合了不同软件对 DDA 和 DIA 两种模式下所得数据的分析结果并进行比较。如图 3 所示,在 DDA 模式下,共定性和定量到 2 894 个和 2 887 个

蛋白;在 DIA 模式下,共定性和定量到 2 661 个和 2 623 个蛋白。以上结果表明 DIA 和 DDA 模式得到的定性定量蛋白数量相当,DIA 对于 DDA 没有明显优势。然而在高等动物的蛋白质组学研究中,DIA 技术的定量能力优于 DDA<sup>[14]</sup>,推测可能是由于本实验以细菌作为研究样本,较为简单,蛋白数目比高等生物少,现有的仪器在 DDA 模式下就可以采集到较为全面的数据。另外,比较了各个软件对 DDA 及 DIA 数据的分析结果。SPE 中得到的 DIA 蛋白数量略少于 DDA 所得结果,这可能是由于 SPE 的分析方法依赖于事先构建好的 DDA 谱图库,而已经构建的 DDA 谱图库不能包含所有肽段信息,导致部分蛋白分析结果丢失<sup>[15]</sup>。因此尝试在 PEAKS 软件中使用搜库的方法对 DDA 和 DIA 的数据同时进行分析,结果表明 DIA 的蛋白鉴定

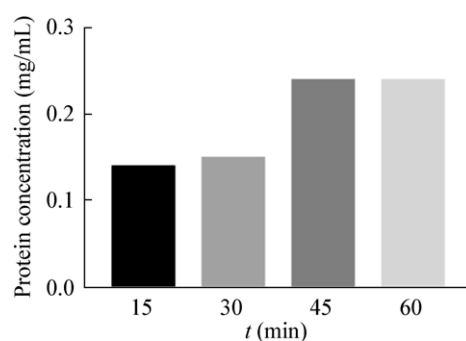


图 2 不同超声时间下鞘脂菌 SJTF-8 全蛋白提取效率比较

Figure 2 Comparison of total protein extraction efficiency of *Sphingobium yanoikuyae* strain SJTF-8 under different breaking time

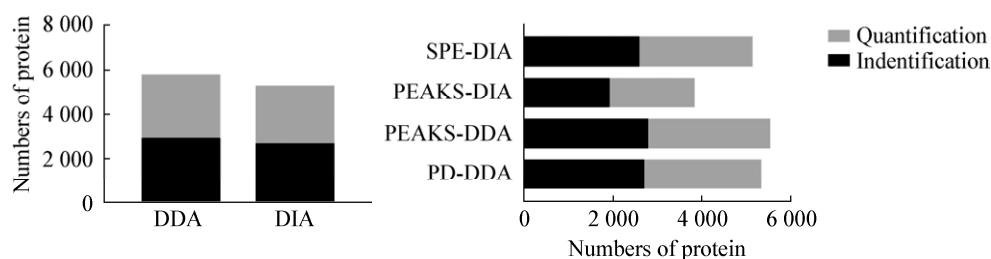


图 3 DDA 与 DIA 定性定量蛋白数目比较

Figure 3 Comparison of identity and quantity protein numbers between DDA and DIA

注: A: DDA 和 DIA 两种质谱模式下采集数据在不同软件中分析结果的整合; B: 4 种软件对 2 种质谱模式下所得数据分别分析的结果。

Note: A: The integration of data collected in different software analysis results under DDA and DIA; B: The respectively analyzing result by four software under two mass spectrometry modes.

数目仍然小于 DDA 中蛋白鉴定数目, 这可能是由于 DIA 二级谱图包含采集窗口内所有子离子信息, 数据量远大于 DDA 的数据量, PEAKS 软件使用 *de novo* 算法结合直接搜库的功能对 DIA 数据进行图谱解析较困难, 从而使得 DIA 谱图解析率达不到 DDA 的水平。

2.2.2 缺失值分析

在对所有数据进行整体分析后, 根据每组中样品的检出度, 将所有定量到的蛋白分成 A 和 B 两组, 如表 2 所示, A 组为非诱导表达而 LB 完全不表达蛋白以及 LB 诱导表达而非完全不表达蛋白, B 组为剔除 A 组蛋白后剩余定量蛋白。A 组蛋白部分样品定性定量的缺失是由于样品本身蛋白表达差异造成, 不适合用来做软件缺失值分析。因此, 仅筛选了 B 组蛋白完成缺失值分析。如表 2 所示, 在 DIA 质谱方法下, 样品的缺失率为 2.38% 和 1.95%, 明显小于 DDA 方法。推测可能是由于在 DDA 模式下, 实时信号采集会根据母离子信号强度歧视性地选择碎裂离子, 对于丰度较低的肽段和相应蛋白很难保证每次实验信号采集的可重复性<sup>[16-17]</sup>。所以, 相比于 DDA 技术, DIA 质谱采集方法得到的数据稳定性更好、可重复性更强、定量的准确性更高。

2.2.3 重现性分析

利用 Spectronaut 软件分析 DIA 数据获得定量蛋白火山图。如图 4 所示, 软件根据蛋白丰度从上到下进行排序后发现, A 组中大多数蛋白为低丰度

蛋白, 并且其中有相当一部分低丰度蛋白仅在非中表达, 说明 DIA 模式对发现鞘脂菌在多环芳烃中低丰度表达的差异蛋白具有一定意义。

为了判断 A 组中低丰度蛋白在 DDA 和 DIA 中的重现性, 我们选取了 PEAKS-DDA 与 SPE-DIA 作为 DDA 和 DIA 结果的代表, 将 SPE-DIA 和 PEAKS-DDA 的 A 组蛋白和对方的所有定量蛋白以及经  $CV \leq 20\%$  筛选后的定量蛋白进行比较。如图 5 所示, SPE-DIA 的 A 组蛋白中有 260 个蛋白在 PEAKS-DDA 中也被鉴定到, 占总蛋白数目的 71.8%。但是经  $CV \leq 20\%$  筛选后重叠蛋白数显著降低, 只有 22 个蛋白在 PEAKS-DDA 中被鉴定到, 约 90% (238 个) 的蛋白在 PEAKS-DDA 定量方式中因为重现性差而被过滤弃用。用同样的方法分析 PEAKS-DDA 的 A 组数据, 在 SPE-DIA 中鉴定到的蛋白数目过滤前后分别为 44 和 30 个, 重叠蛋白数目变化不大。DIA 采集模式的原理是对于所有落在隔离窗口范围内的多肽离子都将被同时碎裂和检测, 因而许多低丰度的蛋白也被系统化、无歧视地一并记录下来, 从而表现出组内蛋白定量变异系数小的特点<sup>[18-19]</sup>。然而在 DDA 中, 低丰度蛋白由于歧视效应较难被可重复性检测, 因而其组内蛋白定量变异系数大, 导致定量准确性差。

2.2.4 DDA 与 DIA 技术方法比较

如表 3 所示, 我们从样品前处理到数据分析及技术性能等多方面比较了 DDA 与 DIA 技术的相同点与不同点。可以发现, 对于 DDA 和 DIA 方法

表 2 两种质谱模式下定量结果缺失值比较

Table 2 Comparison of missing values of quantitative results from DDA and DIA in three different software

| 分析软件<br>Analysis software | 定量蛋白<br>Total quantitative protein | A<br>Protein number | B                      |                       |                         |
|---------------------------|------------------------------------|---------------------|------------------------|-----------------------|-------------------------|
|                           |                                    |                     | 蛋白数量<br>Protein number | 缺失值*<br>Missing value | 缺失率<br>Missing rate (%) |
| PD-DDA                    | 2 645                              | 77                  | 2 568                  | 603                   | 3.91                    |
| PEAKS-DDA                 | 2 757                              | 106                 | 2 652                  | 508                   | 3.19                    |
| PEAKS-DIA                 | 1 919                              | 34                  | 1 885                  | 221                   | 1.95                    |
| SPE-DIA                   | 2 555                              | 362                 | 2 194                  | 313                   | 2.38                    |

注: A: 非诱导表达而 LB 完全不表达蛋白, 以及 LB 诱导表达而非完全不表达蛋白; B: 剔除 A 组的蛋白后剩余定量蛋白; \*: 缺失值根据所有样品中未检出数确定, 缺失率为缺失值与所有样品中蛋白定量总数的比值。

Note: A: Proteins which are expressed only in phenanthrene condition or only in LB condition; B: Residual quantitative proteins after removing proteins from group A; \*: The missing value is determined according to the undetected protein number in all samples, and the missing rate is the ratio of the missing value to the total quantitative amount of protein in all samples.

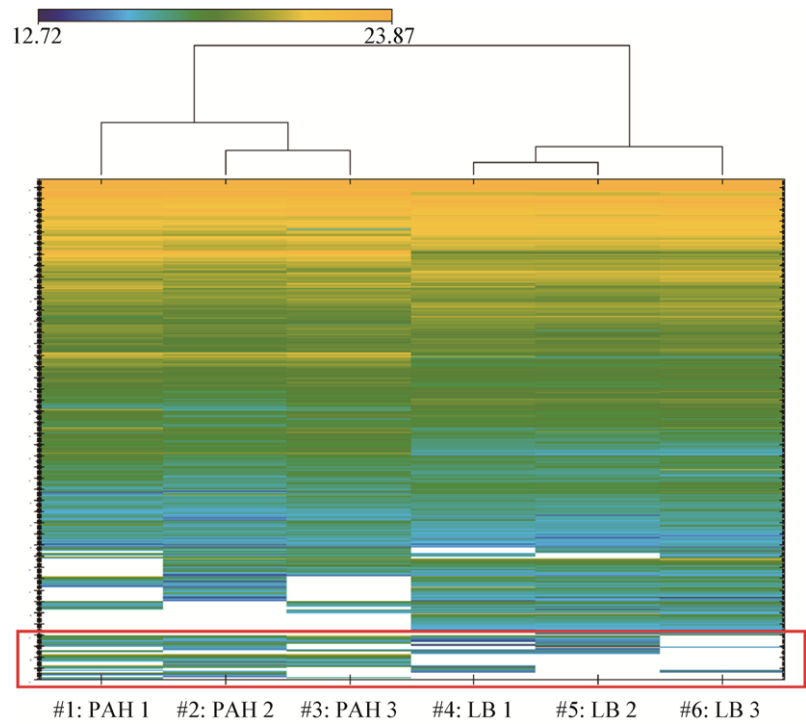


图 4 Spectronaut 中所有定量蛋白火山图  
Figure 4 Volcanic map of all quantitative proteins from Spectronaut

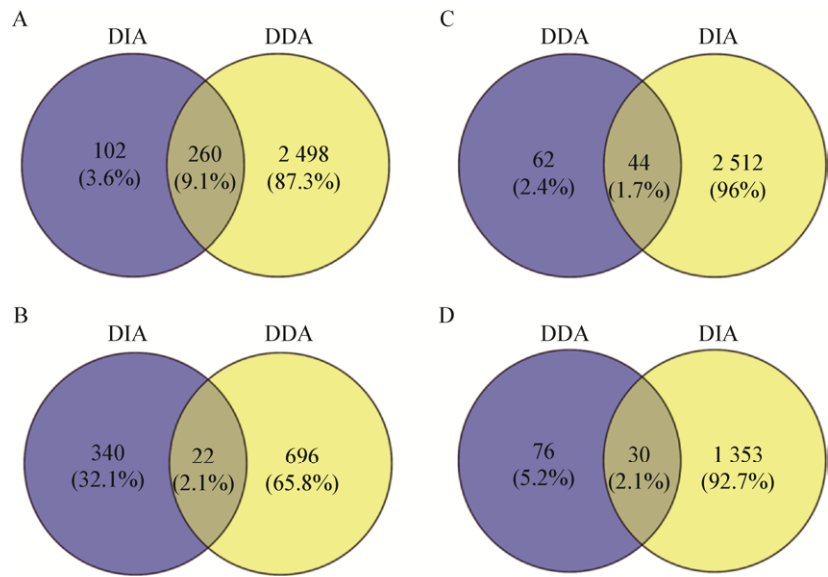


图 5 PEAKS-DDA 和 SPE-DIA 的 A 组蛋白定量结果差异比较  
Figure 5 Difference comparison of quantitative proteins from group A between PEAKS-DDA and SPE-DIA

注:A;SPE-DIA 中 A 组蛋白相对于 PEAKS-DDA 全部定量蛋白的覆盖情况,B;SPE-DIA 中 A 组蛋白相对于 CV≤20%的 PEAKS-DDA 定量蛋白的覆盖情况, C: PEAKS-DDA 中 A 组蛋白相对于 SPE-DIA 全部定量蛋白的覆盖情况, D: PEAKS-DDA 中 A 组蛋白相对于 CV≤20%的 SPE-DIA 定量蛋白的覆盖情况.

Note: A: The coverage of proteins from group A in SPE-DIA relative to all quantitative proteins in PEAKS-DDA; B: The coverage of proteins from group A in SPE-DIA relative to quantitative proteins with CV lower than 20% in PEAKS-DDA; C: The coverage of proteins from group A in PEAKS-DDA relative to all quantitative proteins in SPE-DIA; D: The coverage of proteins from group A in PEAKS-DDA relative to quantitative proteins with CV lower than 20% in SPE-DIA.

而言, 样品前处理与液相色谱方法没有什么差别, 本实验中两种技术也都在 Q-Exactive Plus 仪器上实现。然而根据两种技术的原理不同, DIA 在质谱采集过程中会引入大量肽段并将它们同时碎裂, 从而产生高度混合的 MS2 谱图<sup>[20]</sup>。如此复杂的 MS2 谱图虽然也可以通过传统的基于在线数据库匹配的方法直接分析, 但效果不佳, 因此人们开发了称之为“肽段中心打分”的分析方法。在这项策略中, 采集到的数据集会和一套已知肽段各类参数的数据集进行比对搜索, 最常用的方法就是预先采集一套谱图数据库<sup>[21]</sup>。另外, 由于 DDA 数据采集的随机性和限制性, 其肽段检测的可重复性及定量一致性较差, 而 DIA 模式不涉及到对肽段离子的限制性筛选而获得更完整的信息, 且相对于母离子检测来说, 碎片离子检测的灵敏度更高, 包含的信息也更丰富, 因此 DIA 相较于 DDA 来说更稳健、更灵敏。

2.3 PEAKS 与 Spectronaut 软件对 DIA 数据分析方法比较

对同一批 DIA 数据利用 PEAKS Studio10.0 (Bioinformatics Solutions Inc.) 和 Spectronaut 11.0 (Biognosys AG 2013) 两种搜索工具分别进行分析, 从软件对服务器的要求、处理步骤和效率、定性定量结果等方面比较两种软件的性能。从表 4 分析结果可以看出, 两种软件在分析性能方面各有优缺点。PEAKS 可通过直接搜库的方法对 DIA 数据进行处理, 因此操作步骤较为简单, 省去构建 DDA 实体库的步骤, 前期准备工作较少; 然而由于其融

合 *de novo* 的算法对肽段进行定性分析, 在分析时间上要远长于 SPE 的数据处理时间。同时, PEAKS 软件定量的肽段数目有 23 891, 高于 SPE 软件的 14 641, 但是蛋白定量数目明显低于 SPE 软件, 说明 *de novo* 的方法对肽段的鉴定数量有了较大的提升, 然而对蛋白的定量结果并没有明显帮助。分析其中原因, 可能是由于 DIA 数据较为复杂, 直接进行线上搜库匹配的方法使得谱图匹配效率降低。SPE 通过构建实体库的方法进行 DIA 数据分析, 大大降低了谱图解析难度, 增强了分析效果。除此之外, PEAKS 是利用一级谱图母离子峰面积或者峰强度对蛋白定量, 为了达到定量要求, 一级谱图信噪比要求较高, 增大了低丰度蛋白的定量难度, 导致很多低丰度不能被有效定量<sup>[22]</sup>; 而 SPE 在二级谱图的基础上根据子离子峰面积或者峰强度进行定量, 有利于信噪比的提高和低丰度蛋白的检出, 增强了定量能力<sup>[8]</sup>。

2.4 差异蛋白及生物信息学分析

2.4.1 差异蛋白的筛选

根据图 1 中不同软件对差异蛋白的筛选条件, 分别对 DDA 和 DIA 模式下获得的质谱数据进行差异蛋白分析, 然后将 DDA 在 PD 和 PEAKS 中所得差异蛋白合并以及将 DIA 在 SPE 和 PEKAS 中所得差异蛋白合并; 为了更准确地得到表达差异蛋白, 筛选获得在 DDA 与 DIA 共得到的 580 个差异蛋白进行进一步的生物信息学分析, 差异蛋白列表详见表 5, 其中非诱导的表达下调蛋白有 327 个, 表达上调蛋白有 253 个。

表 3 DDA 与 DIA 技术方法比较

Table 3 Comparison of DDA and DIA method

| Feature      | Category   | DDA  | DIA  |
|--------------|--|--|--|
| Differences  | Principle  | Chooses the largest peaks for acquisition of MS2 spectra and peptide identification  | All ionized compounds are fragmented in a systematic and unbiased fashion                                      |
|              | Data acquisition   | Requires definition of TopN methos, MS2 trigger threshold and dynamic exclusion time | Requires definition of mass range to cover, precursor isolation window width and number of MS2 scans per cycle |
|              | Spectrum library establishment                                     | No requirement   | Generally requires   |
|              | Software   | PD, PEAKS, Mascot, Maxquant  | PEAKS, Spectronaut, OpenSWATH, Skyline   |
|              | Reproducibility  | Low, due to stochastic sampling in DDA   | High, due to peptide-centric scoring analysis  |
| Similarities | Sample treatment method, chromatographic method, mass spectrometer |  |  |



表 4 PEAKS 与 Spectronaut 软件处理 DIA 数据性能比较

Table 4 Performance comparison of PEAKS and Spectronaut software in processing DIA data

| Software | 肽段<br>Peptide | 蛋白<br>Protein | 定性方法<br>Identification    | 定量方法<br>Quantification | 软件安装要求<br>Software installation | 处理步骤<br>Data processing steps | 分析效率<br>Analysis efficiency |
|----------|---------------|---------------|---------------------------|------------------------|---------------------------------|-------------------------------|-----------------------------|
| PEAKS    | 23 891        | 1 921         | <i>de novo</i> +DB search | MS1                    | 16thread                        | Less                          | Slower                      |
| SPE      | 14 641        | 2 595         | Spectral library search   | MS2                    | 16thread                        | More                          | Faster                      |

表 5 在 DDA 与 DIA 共同所得差异蛋白

Table 5 Differentially expressed protein commonly obtained in DDA and DIA

| Down-regulated protein |                    |   | Up-regulated protein |                    |   |
|------------------------|--------------------|---|----------------------|--------------------|---|
| No.                    | Protein accessions | Protein names   | No.                  | Protein accessions | Protein names   |
| 1                      | A0A085K0S9         | L-ectoine synthase  | 1                    | A0A085K9P4         | Formyltetrahydrofolate deformylase  |
| 2                      | A0A084ENS0         | 50S ribosomal protein L32   | 2                    | A0A085KA57         | Glucoamylase (Glycoside hydrolase family 15 protein)  |
| 3                      | A0A085KB35         | 30S ribosomal protein S20   | 3                    | A0A3G2UT35         | Organic hydroperoxide resistance protein  |
| 4                      | A0A085K2P5         | Sec-independent protein translocase protein TatC                                    | 4                    | A0A085KAK8         | UPF0301 protein AX777_21150   |
| 5                      | A0A085K6N1         | Aromatic amino acid aminotransferase (Aspartate/tyrosine/aromatic aminotransferase) | 5                    | A0A085JYY1         | Tetratricopeptide repeat protein  |
| 6                      | A0A084EPD8         | 50S ribosomal protein L15   | 6                    | A0A085K5K6         | TonB-dependent receptor (TonB-dependent siderophore receptor)   |
| 7                      | A0A084EUR9         | Nucleoside diphosphate kinase (NDK) (NDP kinase)                                    | 7                    | A0A3G2UKT1         | IS6 family transposase  |
| 8                      | A0A084ECG9         | 50S ribosomal protein L35   | 8                    | A0A085K649         | Signal peptidase I  |
| 9                      | A0A3G2UTA2         | Adenine phosphoribosyltransferase (APRT)  | 9                    | A0A3G2UTD7         | Polysaccharide export protein   |
| 10                     | A0A085K7P9         | Alanine dehydrogenase   | 10                   | A0A085K5I6         | NAD-dependent succinate-semialdehyde dehydrogenase (Succinate-semialdehyde dehydrogenase)   |
| 11                     | A0A3G2UQQ0         | Peptide MFS transporter   | 11                   | A0A085K062         | TonB-dependent receptor   |
| 12                     | A0A084EHR1         | 30S ribosomal protein S21   | 12                   | A0A084EUH6         | Chemotaxis protein CheY (DNA-binding response regulator) (Response regulator of the LytR/AlgR family protein) (Two-component system response regulator)                         |
| 13                     | A0A085K0T0         | Ectoine hydroxylase   | 13                   | A0A085JZ02         | Uncharacterized protein   |
| 14                     | A0A3G2V2E7         | Acylase   | 14                   | A0A085K9E4         | Histidine kinase (Response regulator)   |
| 15                     | A0A084ELD5         | Lipoyl synthase   | 15                   | A0A085KA86         | Alpha-D-glucose phosphate-specific phosphoglucomutase (Phosphoglucomutase)  |
| 16                     | A0A085K0I3         | 50S ribosomal protein L21   | 16                   | A0A085K729         | TonB-dependent receptor (TonB-dependent siderophore receptor)   |
| 17                     | A0A085K2A3         | 30S ribosomal protein S10   | 17                   | A0A085K9I2         | Class I SAM-dependent methyltransferase (Cyclopropane-fatty-acyl-phospholipid synthase) (Methyltransferase, cyclopropane fatty acid synthase) (SAM-dependent methyltransferase) |
| 18                     | A0A085K299         | 30S ribosomal protein S12   | 18                   | A0A085K7B1         | Nitrite/sulfite reductase (Sulfite reductase)   |
| 19                     | A0A3G2UPG2         | Oleate hydratase  | 19                   | A0A085KB08         | 3-dehydroquinate dehydratase (3-dehydroquinase)   |
| 20                     | A0A084ET38         | 30S ribosomal protein S4  | 20                   | A0A085JZI0         | Membrane protein (SPFH/Band 7/PHB domain protein)   |
| 21                     | A0A085K1S4         | 30S ribosomal protein S15   | 21                   | A0A085K679         | DUF853 family protein   |
| 22                     | A0A085K2G0         | Peptidase S10   | 22                   | A0A3G2US14         | Copper resistance protein B   |
| 23                     | A0A084E3I3         | Arsenate reductase  | 23                   | A0A085K6I1         | Uncharacterized protein   |
| 24                     | A0A085K6N3         | Homogentisate 1,2-dioxygenase (HGDO)  | 24                   | A0A3G2UPU6         | Polysaccharide biosynthesis tyrosine autokinase   |

(待续)

(续表 5)

|    |            |   |    |            |  |
|----|------------|---|----|------------|--|
| 25 | A0A085K6X3 | Phosphogluconate dehydratase  | 25 | A0A085K7F0 | ABC transporter substrate-binding protein (Lipoprotein-releasing ABC transporter permease subunit) |
| 26 | A0A084EBB1 | HAD family hydrolase  | 26 | A0A084ELN5 | Peptidoglycan-associated protein   |
| 27 | A0A084EFC1 | HU family DNA-binding protein (Integration host factor) (Transcriptional regulator)   | 27 | A0A085K6Y5 | 3-hydroxyacyl-CoA dehydrogenase  |
| 28 | I0IW05     | 30S ribosomal protein S8  | 28 | A0A3G2UPQ0 | NADH:flavin oxidoreductase/NADH oxidase  |
| 29 | A0A085K012 | 50S ribosomal protein L27   | 29 | A0A3G2URM5 | Aldehyde dehydrogenase family protein  |
| 30 | A0A085KA25 | Alpha-L-fucosidase  | 30 | A0A085KB61 | Beta-glucosidase (Beta-glucosidase BglX)   |
| 31 | A0A085K4Y1 | Amino acid dehydrogenase (Glu/Leu/Phe/Val dehydrogenase)  | 31 | A0A085KA56 | Trehalose-6-phosphate synthase   |
| 32 | A0A085KAT1 | 50S ribosomal protein L28   | 32 | A0A085K311 | Cytochrome C (Cytochrome c family protein)   |
| 33 | A0A084EUN9 | Dihydroxy-acid dehydratase  | 33 | A0A085K7P8 | AsnC family transcriptional regulator (Lrp/AsnC family transcriptional regulator)                  |
| 34 | A0A085K8U7 | Helix-turn-helix transcriptional regulator (Peptidase S24)  | 34 | A0A3G2UTB7 | UDP-glucose 6-dehydrogenase  |
| 35 | A0A085K8D1 | TIGR02300 family protein  | 35 | A0A085K1H2 | TonB-dependent receptor  |
| 36 | A0A085K9X4 | Urocanate hydratase (Urocanase)   | 36 | A0A085KAR7 | Membrane protein   |
| 37 | A0A084E4U8 | DNA-binding protein (Heat shock protein HspQ) (Hemimethylated DNA binding domain-containing protein)  | 37 | A0A084ELS5 | AAA family ATPase (Cell division protein FtsH)   |
| 38 | A0A084EL94 | Biotin synthase   | 38 | A0A085K801 | Cytochrome C (Cytochrome c1)   |
| 39 | A0A085K2A6 | 50S ribosomal protein L23   | 39 | A0A3G2UMU2 | IclR family transcriptional regulator  |
| 40 | A0A085K6N2 | 4-hydroxyphenylpyruvate dioxygenase   | 40 | A0A085K5D3 | Outer-membrane lipoprotein carrier protein   |
| 41 | A0A085K7W6 | Flagellin   | 41 | A0A085KBB1 | DUF4136 domain-containing protein  |
| 42 | A0A085K5E1 | Uncharacterized protein   | 42 | A0A085K671 | Opacity protein  |
| 43 | A0A085KA36 | Glycerol kinase   | 43 | A0A3G2UQ29 | Glycosyltransferase family 2 protein   |
| 44 | A0A3G2ULM1 | Aromatic ring-hydroxylating dioxygenase subunit alpha   | 44 | A0A085K1M7 | Glutathione S-transferase (Glutathione S-transferase family protein)                               |
| 45 | A0A085K6N9 | Corrinoid adenosyltransferase (Cob(II)alamin adenosyltransferase)   | 45 | A0A3G2UQ32 | TrbI/VirB10 family protein   |
| 46 | A0A085K2T8 | Methyltransferase   | 46 | A0A084ES41 | DUF1508 domain-containing protein  |
| 47 | A0A3G2UPV1 | L-2,4-diaminobutyric acid acetyltransferase (DABA acetyltransferase)  | 47 | A0A3G2UQM6 | NAD(P)-dependent oxidoreductase  |
| 48 | A0A3G2UX96 | Phosphofructokinase   | 48 | A0A085K603 | Fumarate hydratase class II (Fumarase C)   |
| 49 | A0A085K4Q1 | ArsR family transcriptional regulator (Putative ArsR family transcriptional regulator) (Transcriptional regulator)                              | 49 | A0A085K235 | Membrane protein   |
| 50 | A0A085K526 | Cytochrome P450   | 50 | A0A3G2UM71 | DUF4142 domain-containing protein  |
| 51 | A0A085K6V0 | Glycoside hydrolase family 105 protein (Glycosyl hydrolase family 88)   | 51 | A0A0J9D3E5 | Uncharacterized protein  |
| 52 | A0A085K9A8 | Beta-lactamase (Serine hydrolase)   | 52 | A0A085K681 | Cysteine synthase  |
| 53 | A0A084EDE6 | Acetolactate synthase (Thiamine pyrophosphate-binding protein) (Thiamine pyrophosphate-dependent enzyme, possible carboxylase or decarboxylase) | 53 | A0A3G2UR44 | Dicarboxylate/amino acid:cation symporter  |
| 54 | A0A085KB48 | Aminobenzoate synthetase (Aminotransferase) (Para-aminobenzoate synthase, component I)  | 54 | A0A085K478 | Opacity protein  |
| 55 | A0A084ECJ4 | FAD-dependent oxidoreductase (NAD(P)/FAD-dependent oxidoreductase)  | 55 | A0A084ECW6 | CoA-binding protein (Putative CoA-binding protein)   |
| 56 | A0A085JZ36 | PspC domain-containing protein (Stress-responsive transcriptional regulator)  | 56 | A0A085K047 | Uncharacterized protein  |
| 57 | A0A085K239 | Uncharacterized protein   | 57 | A0A085K8X5 | Glutamate synthase (Glutamate synthase large subunit)  |

(待续)

(续表 5)

|    |            |  |    |            |   |
|----|------------|--|----|------------|---|
| 58 | A0A085K242 | Pirin (Pirin family protein)   | 58 | A0A085KAH0 | Acetyl-CoA C-acetyltransferase (Acetyl-CoA acetyltransferase)                               |
| 59 | A0A085K5A7 | UPF0260 protein EBF16_22055  | 59 | A0A3G2UMR2 | Uncharacterized protein   |
| 60 | A0A085K5L0 | Thioesterase (Thioesterase family protein)   | 60 | A0A085K4Y8 | Signal protein (TonB-dependent receptor)  |
| 61 | A0A085K5Y5 | Uncharacterized protein  | 61 | A0A3G2UPP1 | 4-oxalocrotonate decarboxylase  |
| 62 | A0A085K6F8 | Diguanylate cyclase (GGDEF domain-containing protein)                                  | 62 | A0A3G2ULZ9 | Uncharacterized protein   |
| 63 | A0A085K6P7 | DUF721 domain-containing protein   | 63 | A0A3G2V2Q8 | PadR family transcriptional regulator   |
| 64 | A0A085K6X7 | Uncharacterized protein  | 64 | A0A085K2K3 | Metal/formaldehyde-sensitive transcriptional repressor (Transcriptional regulator)          |
| 65 | A0A085K809 | DUF3240 domain-containing protein  | 65 | A0A085K8S2 | Glycerol kinase   |
| 66 | A0A085KA91 | Polysaccharide biosynthesis protein GumN (TraB/GumN family protein)                    | 66 | A0A085K494 | Uncharacterized protein   |
| 67 | A0A085KBB9 | DUF2188 domain-containing protein  | 67 | A0A085K7I7 | EmrA/EmrK family multidrug efflux transporter periplasmic adaptor subunit (Hemolysin D)     |
| 68 | A0A3G2UKF6 | Polyisoprenoid-binding protein   | 68 | A0A085JZ61 | ABC transporter permease (FtsX-like permease family protein)                                |
| 69 | A0A3G2UNN8 | Antitoxin  | 69 | A0A3G2UN54 | AarF/ABC1/UbiB kinase family protein  |
| 70 | A0A3G2UTE3 | DUF4880 domain-containing protein  | 70 | A0A085K253 | Penicillin-binding protein activator LpoB (Protein involved in formation of curli polymers) |
| 71 | A0A3G2V109 | Uncharacterized protein  | 71 | A0A3G2UTG9 | SGNH_hydro domain-containing protein  |
| 72 | A0A3G2V1E7 | DUF2779 domain-containing protein  | 72 | A0A085K800 | Cytochrome b  |
| 73 | A0A3G2V1F3 | Uncharacterized protein  | 73 | A0A3G2UZ77 | LLM class flavin-dependent oxidoreductase   |
| 74 | A0A085K7V9 | Flagellar biosynthesis anti-sigma factor FlgM  | 74 | A0A085KAY9 | Membrane protein  |
| 75 | A0A085KAY2 | Chemotaxis protein CheY (Response regulator) (Two-component system response regulator) | 75 | A0A085K766 | Glycosyltransferase family 2 protein (Histidine kinase)                                     |
| 76 | A0A085K5C1 | Putative pyruvate, phosphate dikinase regulatory protein (PPDK regulatory protein)     | 76 | A0A085K485 | Catalase  |
| 77 | A0A3G2UUG6 | Coproporphyrinogen-III oxidase   | 77 | A0A177JY28 | Uncharacterized protein   |
| 78 | A0A3G2UTF0 | Type II toxin-antitoxin system ParD family antitoxin                                   | 78 | A0A085K1E8 | OsmC family peroxiredoxin (Peroxiredoxin)   |
| 79 | A0A3G2UKT7 | Cytochrome b   | 79 | A0A085KA00 | Uncharacterized protein   |
| 80 | A0A084EII0 | Uncharacterized protein  | 80 | A0A085K8S3 | Catalase  |
| 81 | A0A3G2UXD4 | ROK family protein   | 81 | A0A2D1R5E7 | Uncharacterized protein   |
| 82 | A0A3G2UKI7 | Pirin family protein   | 82 | A0A084EJW0 | DedA family protein (Membrane protein) (Putative membrane-associated protein)               |
| 83 | A0A085K7A8 | DUF885 domain-containing protein (Tat pathway signal protein)                          | 83 | A0A085K3D5 | TonB-dependent receptor   |
| 84 | A0A084EI05 | DUF4170 domain-containing protein  | 84 | A0A085K8R8 | Phasin (Phasin family protein)  |
| 85 | A0A085JZY0 | Probable glycine dehydrogenase (decarboxylating) subunit 1                             | 85 | A0A084EUC7 | LemA family protein (Membrane protein)  |
| 86 | A0A3G2US79 | Uncharacterized protein  | 86 | A0A3G2V0F0 | Uncharacterized protein   |
| 87 | A0A085K2U3 | Pilus assembly protein CpaE  | 87 | A0A3G2UL12 | Uncharacterized protein   |
| 88 | A0A085K9S7 | Peptidase M28 (Peptidase M28 family protein)   | 88 | A0A085K0K3 | Copper resistance system multicopper oxidase (Copper-binding protein)                       |
| 89 | A0A085K7J2 | 30S ribosomal protein S16  | 89 | A0A3G2UMU8 | Murein L,D-transpeptidase   |
| 90 | A0A085KAU4 | 50S ribosomal protein L10  | 90 | A0A3G2UTB5 | Efflux transporter outer membrane subunit   |
| 91 | I0IW02     | 30S ribosomal protein S17  | 91 | A0A085K314 | Membrane protein (OmpA family protein)  |
| 92 | I0IW03     | 50S ribosomal protein L24  | 92 | A0A085K9N5 | Dihydroxy-acid dehydratase (DAD)  |
| 93 | A0A084ERX6 | GTPase Obg   | 93 | A0A084EMQ0 | DUF1134 domain-containing protein   |
| 94 | A0A085K558 | Peptidase M61  | 94 | A0A3G2UV24 | Autotransporter domain-containing protein   |
| 95 | A0A3G2ULC1 | TIGR01244 family phosphatase   | 95 | A0A085K8X6 | Dihydropyrimidine dehydrogenase (NAD(P)-dependent oxidoreductase)                           |

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| 96  | I0IVZ9     | 30S ribosomal protein S19  | 96  | A0A3G2USP8 | SDR family oxidoreductase   |
| 97  | A0A085K2B2 | 50S ribosomal protein L29  | 97  | A0A085K651 | Uncharacterized protein   |
| 98  | A0A085K983 | Aminotransferase   | 98  | A0A084EU87 | DUF4136 domain-containing protein (Lipoprotein transmembrane)   |
| 99  | A0A085K918 | Dual-specificity RNA methyltransferase RlmN  | 99  | A0A3G2USM1 | Quinone oxidoreductase  |
| 100 | A0A085K375 | 50S ribosomal protein L31  | 100 | A0A3G2UM34 | Uncharacterized protein   |
| 101 | A0A177JXN1 | 2-oxoisovalerate dehydrogenase (Alpha-ketoacid dehydrogenase subunit beta)                         | 101 | A0A085K553 | DUF4139 domain-containing protein   |
| 102 | A0A085K2E4 | Heme chaperone HemW  | 102 | A0A3G2UTA9 | RraA family protein   |
| 103 | A0A085JZ30 | M13 family peptidase (Peptidase M13)   | 103 | A0A085K655 | Aldolase (Class II aldolase/adducin family protein)   |
| 104 | A0A085K5Z7 | Crp/Fnr family transcriptional regulator (cAMP-binding protein-catabolite transcription activator) | 104 | A0A085KBB6 | UPF0145 protein A6768_25245   |
| 105 | A0A085K017 | 50S ribosomal protein L1   | 105 | A0A085K3P1 | DNA starvation/stationary phase protection protein (DNA-binding protein) (Putative low temperature-induced protein) |
| 106 | A0A085K960 | Zinc-finger domain-containing protein  | 106 | A0A3G2UPK4 | 2-oxo-hepta-3-ene-1,7-dioic acid hydratase  |
| 107 | I0IW00     | 50S ribosomal protein L22  | 107 | A0A3G2URD7 | Polyisoprenoid-binding protein  |
| 108 | A0A3G2UPQ4 | Aromatic ring-hydroxylating dioxygenase subunit alpha  | 108 | A0A085K8R3 | Aromatic ring-opening dioxygenase LigA (Dioxygenase)  |
| 109 | A0A085K9P9 | 2-nitropropane dioxygenase (Nitronate monooxygenase)   | 109 | A0A3G2UYK9 | Antibiotic biosynthesis monooxygenase   |
| 110 | A0A085K9G3 | Dihydrofolate reductase  | 110 | A0A3G2UK76 | Alcohol dehydrogenase AdhP  |
| 111 | A0A085K949 | TonB-dependent receptor  | 111 | A0A085K790 | NAD(P)H dehydrogenase (quinone)   |
| 112 | A0A085K2A5 | 50S ribosomal protein L4   | 112 | A0A3G2UQL4 | TonB-dependent receptor   |
| 113 | A0A085K2G6 | Oxidoreductase (SDR family oxidoreductase)   | 113 | A0A3G2UMG6 | 3-phenylpropionate/cinnamic acid dioxygenase subunit beta   |
| 114 | A0A085K2A7 | 50S ribosomal protein L2   | 114 | A0A085K9E7 | Aldehyde oxidase (Aldo/keto reductase)  |
| 115 | A0A084EPD6 | Adenylate kinase (AK)  | 115 | A0A085K991 | D-3-phosphoglycerate dehydrogenase  |
| 116 | A0A084EHB6 | Transcription antitermination protein NusB (Antitermination factor NusB)                           | 116 | A0A3G2ULT9 | Benzene 1,2-dioxygenase   |
| 117 | A0A085K6Z9 | Uncharacterized protein  | 117 | A0A084ENV1 | DUF3617 domain-containing protein   |
| 118 | A0A085K705 | 50S ribosomal protein L9   | 118 | A0A085K015 | Cation:proton antiport protein (Kef family K(+) transporter)  |
| 119 | A0A085K5B5 | PTS IIA-like nitrogen-regulatory protein PtsN (PTS lactose transporter subunit IIC)                | 119 | A0A3G2ULR1 | NAD(P)-dependent alcohol dehydrogenase  |
| 120 | A0A085K5C2 | Uroporphyrinogen decarboxylase (UPD)   | 120 | A0A3G2UMC1 | Aromatic ring-hydroxylating dioxygenase subunit alpha   |
| 121 | A0A085K139 | DEAD/DEAH box helicase   | 121 | A0A3G2ULV0 | Acetaldehyde dehydrogenase  |
| 122 | A0A084EQ82 | CarD family transcriptional regulator  | 122 | A0A3G2UPW6 | Type 1 glutamine amidotransferase   |
| 123 | A0A085K9A1 | Pyridoxine/pyridoxamine 5'-phosphate oxidase   | 123 | A0A3G2ULX1 | Aromatic-ring-hydroxylating dioxygenase subunit beta  |
| 124 | A0A085K719 | Acyl carrier protein (ACP)   | 124 | A0A3G2UPR4 | DUF3597 family protein  |
| 125 | A0A085K2A0 | 30S ribosomal protein S7   | 125 | A0A3G2UNJ0 | Tartrate dehydrogenase  |
| 126 | A0A085K6R7 | M20/M25/M40 family metallo-hydrolase (Peptidase M28)   | 126 | A0A3G2ULK5 | Amidohydrolase  |
| 127 | A0A3G2UN82 | Acetyl/propionyl/methylcrotonyl-CoA carboxylase subunit alpha                                      | 127 | A0A084EII2 | Pirin (Pirin family protein) (Pirin-related protein)  |
| 128 | A0A085K2G3 | Amidohydrolase (Peptidase M20)   | 128 | A0A3G2UM30 | FAD-binding oxidoreductase  |
| 129 | A0A085KAK0 | Peptidase S9 (S9 family peptidase)   | 129 | A0A3G2UTT5 | Aldehyde dehydrogenase family protein   |
| 130 | A0A085K0V3 | Aconitate hydratase (Bifunctional aconitate hydratase)   | 130 | A0A085KB57 | Cell envelope biogenesis protein OmpA (OmpA family protein)   |
| 131 | A0A085K7Y6 | Acetylornithine aminotransferase (ACOAT)   | 131 | A0A3G2UMI7 | Aldehyde dehydrogenase family protein   |

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| 132 | I0IW06     | 50S ribosomal protein L18  | 132 | A0A3G2UPW2 | 3-(Cis-5,6-dihydroxycyclohexa-1,3-dien-1-yl)propanoate dehydrogenase   |
| 133 | A0A085K2C2 | 50S ribosomal protein L30  | 133 | A0A3G2UTS5 | Alpha/beta fold hydrolase  |
| 134 | A0A084ELC0 | Adenosylhomocysteinase (S-adenosyl-L-homocysteine hydrolase) (AdoHcyase)   | 134 | A0A3G2UQK3 | Uncharacterized protein  |
| 135 | A0A085K980 | Nicotinate-nucleotide-dimethylbenzimidazole phosphoribosyltransferase  | 135 | A0A3G2UM69 | 1,6-dihydroxycyclohexa-2,4-diene-1-carboxylate dehydrogenase   |
| 136 | I0IW04     | 30S ribosomal protein S14  | 136 | A0A3G2V6Z5 | Hydrolase  |
| 137 | A0A3G2UL71 | Sulfonamide-resistant dihydropteroate synthase Sul4  | 137 | A0A3G2UJT5 | Aldehyde dehydrogenase family protein  |
| 138 | A0A085JZ31 | ATP-dependent dethiobiotin synthetase BioD   | 138 | A0A085K4C7 | DUF4142 domain-containing protein (Membrane protein)   |
| 139 | A0A177JYJ0 | Methionine aminopeptidase (MAP) (MetAP)  | 139 | A0A3G2ULQ7 | Flavin prenyltransferase UbiX  |
| 140 | A0A084ELS6 | Methyltransferase (Methyltransferase domain-containing protein) (Phospholipid N-methyltransferase)                             | 140 | A0A3G2UN96 | UbiD family decarboxylase  |
| 141 | A0A085K647 | Pyridoxine 5'-phosphate synthase (PNP synthase)  | 141 | A0A3G2ULI3 | 4-hydroxy-2-oxovalerate aldolase (HOA) (EC 4.1.3.39) (4-hydroxy-2-keto-pentanoic acid aldolase) (4-hydroxy-2-oxopentanoate aldolase) |
| 142 | A0A3G2URH6 | Nucleoid-associated protein EBF16_06280  | 142 | A0A3G2ULF3 | 2-dehydro-3-deoxyglucarate aldolase  |
| 143 | A0A085K8V5 | Alkylphosphonate utilization protein (PhnA protein) (Putative Zn-ribbon-containing protein involved in phosphonate metabolism) | 143 | A0A3G2UNE0 | Aldolase   |
| 144 | A0A084EPP2 | 30S ribosomal protein S11  | 144 | A0A3G2UPT0 | FAD-binding oxidoreductase   |
| 145 | A0A3G2URN1 | Peptidase  | 145 | A0A3G2USX6 | 2-dehydropantoate 2-reductase  |
| 146 | A0A085K5C5 | ATPase   | 146 | A0A3G2UMN4 | Aromatic ring-hydroxylating dioxygenase subunit alpha  |
| 147 | A0A085K952 | Ribonucleoside-diphosphate reductase   | 147 | A0A084EKV2 | DUF541 domain-containing protein (Membrane protein)  |
| 148 | A0A085K5E7 | HU family DNA-binding protein (Integration host factor)  | 148 | A0A3G2UNB8 | 2-hydroxymuconic semialdehyde dehydrogenase  |
| 149 | A0A085K3P6 | Hsp70 family protein (Molecular chaperone Hsp70)   | 149 | A0A3G2UQV1 | DUF2312 domain-containing protein  |
| 150 | A0A085KBT0 | Aminopeptidase   | 150 | A0A3G2UTQ5 | Cytochrome c   |
| 151 | A0A0J9CTP8 | Fructose-1,6-bisphosphatase class 1 (FBPase class 1)   | 151 | A0A3G2UMK8 | 2-keto-4-pentenoate hydratase  |
| 152 | A0A085K244 | Pirin (Pirin family protein)   | 152 | A0A3G2UPJ2 | Dihydrodipicolinate synthase family protein  |
| 153 | A0A085K7F4 | DNA polymerase III subunit alpha   | 153 | A0A3G2ULE1 | Pyruvate, phosphate dikinase   |
| 154 | A0A085K9W1 | DUF885 domain-containing protein   | 154 | A0A3G2UU56 | Uncharacterized protein  |
| 155 | A0A3G2V0D2 | LacI family DNA-binding transcriptional regulator  | 155 | A0A084ENF4 | Arabinose ABC transporter permease (MFS transporter)   |
| 156 | A0A085K5G2 | CAP10 domain-containing protein  | 156 | A0A3G2ULK2 | Glutathione transferase GstA   |
| 157 | A0A291N6W4 | Chemotaxis protein   | 157 | A0A3G2UT02 | Anthranilate 1,2-dioxygenase   |
| 158 | A0A085K3L0 | tRNA-specific 2-thiouridylase MnmA   | 158 | A0A3G2UTR6 | Rieske (2Fe-2S) protein  |
| 159 | A0A3G2UND1 | tRNA-2-methylthio-N(6)-dimethylallyladenosine synthase   | 159 | A0A085K990 | Phosphoserine transaminase   |
| 160 | A0A084ERW5 | 50S ribosomal protein L11  | 160 | A0A3G2UM44 | Non-heme iron oxygenase ferredoxin subunit   |
| 161 | A0A085K530 | Methylmalonate-semialdehyde dehydrogenase (Methylmalonate-semialdehyde dehydrogenase (CoA acylating))                          | 161 | A0A3G2UM20 | LLM class flavin-dependent oxidoreductase  |
| 162 | A0A085KB40 | Deoxyuridine 5'-triphosphate nucleotidohydrolase (dUTPase)   | 162 | A0A3G2V0M7 | Uncharacterized protein  |

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| 163 | A0A3G2UN00 | Methylenetetrahydrofolate-tRNA-(uracil-5-)-methyltransferase TrmFO   | 163 | A0A3G2UPL7 | TonB-dependent receptor  |
| 164 | A0A085K328 | Alkaline phosphatase   | 164 | A0A3G2ULG5 | NAD-dependent succinate-semialdehyde dehydrogenase   |
| 165 | A0A085K405 | 30S ribosomal protein S9   | 165 | A0A3G2UPM8 | Rieske (2Fe-2S) protein  |
| 166 | A0A3G2V0E0 | Imidazolonepropionase  | 166 | A0A3G2UPU4 | Anthranilate 1,2-dioxygenase   |
| 167 | A0A3G2V1D5 | SAM-dependent methyltransferase  | 167 | A0A3G2USY5 | Alpha/beta fold hydrolase  |
| 168 | A0A3G2UM87 | MBL fold metallo-hydrolase   | 168 | A0A085K4U8 | Aldehyde dehydrogenase (Aldehyde dehydrogenase family protein)                             |
| 169 | A0A085K6Q5 | Phosphoribosylamine-glycine ligase   | 169 | A0A3G2UMJ7 | Benzoate 1,2-dioxygenase large subunit   |
| 170 | A0A085K3G9 | Adenosine kinase (Carbohydrate kinase)   | 170 | A0A3G2UPV3 | Catechol 2,3-dioxygenase   |
| 171 | A0A085K2B9 | 50S ribosomal protein L6   | 171 | A0A3G2ULH1 | 4-hydroxythreonine-4-phosphate dehydrogenase   |
| 172 | A0A3G2UWN2 | Glycerol-3-phosphate dehydrogenase   | 172 | A0A3G2USZ4 | TonB-dependent receptor  |
| 173 | A0A085K2A2 | Elongation factor Tu (EF-Tu)   | 173 | A0A3G2UNA7 | Biphenyl-2,3-diol 1,2-dioxygenase  |
| 174 | A0A085K7E0 | M28 family peptidase (Peptidase M28)   | 174 | A0A085K8Q8 | Membrane protein   |
| 175 | A0A085K6R8 | Insulinase family protein (Peptidase M16)  | 175 | A0A0J9D4V2 | Cytochrome c oxidase subunit 1   |
| 176 | A0A085K9X2 | Histidine ammonia-lyase (Histidase)  | 176 | A0A085K9C3 | Glycosyltransferase  |
| 177 | A0A085K539 | Putative Zn-dependent protease-like protein (TldD protein) (TldD/PmbA family protein)  | 177 | A0A085K2F7 | Isocitrate lyase   |
| 178 | A0A085KBE2 | Peptidase S10  | 178 | A0A085JZZ6 | Heme exporter protein B  |
| 179 | A0A085K6C8 | 3-hydroxybutyryl-CoA dehydrogenase   | 179 | A0A3G2V2U1 | Acyl-CoA dehydrogenase   |
| 180 | A0A085K8P2 | ArsC family reductase (ArsC family transcriptional regulator) (Arsenate reductase) (Spx/MgsR family transcriptional regulator)           | 180 | A0A3G2UT58 | Enoyl-[acyl-carrier-protein] reductase [NADH]  |
| 181 | A0A3G2V5X6 | Signal recognition particle receptor FtsY (SRP receptor)   | 181 | A0A3G2UT82 | Type 1 glutamine amidotransferase domain-containing protein                                |
| 182 | A0A085K2B0 | 30S ribosomal protein S3   | 182 | A0A3G2UP32 | 1,4-alpha-glucan branching enzyme GlgB   |
| 183 | A0A085KBB7 | Probable transcriptional regulatory protein A6768_25240  | 183 | A0A085K956 | Ammonium transporter   |
| 184 | A0A084EU33 | Translation initiation factor IF-3   | 184 | A0A3G2USV3 | Acyl-CoA synthetase  |
| 185 | A0A085K950 | Fumarate hydratase class I   | 185 | A0A3G2URQ1 | Xanthine dehydrogenase family protein subunit M  |
| 186 | A0A084ENX3 | Beta sliding clamp   | 186 | A0A085K8R0 | ATP-dependent RNA helicase   |
| 187 | A0A085K236 | Recombinase RecJ (Single-stranded-DNA-specific exonuclease RecJ)   | 187 | A0A085K4Z3 | Glycoside hydrolase family 43 (HlyD family efflux transporter periplasmic adaptor subunit) |
| 188 | A0A084ETU1 | UPF0335 protein AX777_20220  | 188 | A0A085K3L6 | K(+)-insensitive pyrophosphate-energized proton pump                                       |
| 189 | A0A085K0S8 | Diaminobutyrate-2-oxoglutarate transaminase  | 189 | A0A085KAF0 | AarF/ABC1/UbiB kinase family protein (Ubiquinone biosynthesis protein)                     |
| 190 | A0A084EHA2 | Translation initiation factor IF-1   | 190 | A0A3G2UYC2 | Lactoylglutathione lyase   |
| 191 | A0A085K3N8 | 50S ribosomal protein L33  | 191 | A0A085K6P9 | NAD(+) diphosphatase   |
| 192 | A0A085K793 | Chemotaxis protein CheY (DNA-binding response regulator) (DNA-binding response regulator CtrA) (Two-component system response regulator) | 192 | A0A3G2UY88 | SDR family oxidoreductase  |
| 193 | A0A085K2N6 | 4-hydroxy-3-methylbut-2-enyl diphosphate reductase (HMBPP reductase)   | 193 | A0A085JZT0 | Glutathione S-transferase (Glutathione S-transferase family protein)                       |
| 194 | A0A3G2UV55 | M20/M25/M40 family metallo-hydrolase   | 194 | A0A085K951 | Glutathione S-transferase (Glutathione S-transferase family protein)                       |
| 195 | A0A084EPE0 | 30S ribosomal protein S5   | 195 | A0A085K9D0 | Acyltransferase  |
| 196 | A0A085K9R7 | tRNA-dihydrouridine synthase   | 196 | A0A085K7C0 | UrcA family protein  |
| 197 | A0A085K5Z4 | Universal stress protein (Universal stress protein UspA)   | 197 | A0A3G2UT41 | Uncharacterized protein  |

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|-----|------------|---|-----|------------|--|
| 198 | A0A085K0Z3 | DUF2322 family protein  | 198 | A0A085JZ29 | DUF4168 domain-containing protein                                    |
| 199 | A0A085K986 | 2,3,4,5-tetrahydropyridine-2,6-dicarboxylate N-succinyltransferase  | 199 | A0A3G2USH6 | YIP1 family protein  |
| 200 | A0A084EQT5 | Ornithine carbamoyltransferase (OTCase)   | 200 | A0A3G2UYE6 | DUF4142 domain-containing protein                                    |
| 201 | A0A084EI43 | Elongation factor P (EF-P)  | 201 | A0A085KA97 | Uncharacterized protein  |
| 202 | A0A085JZ22 | DNA gyrase inhibitor YacG   | 202 | A0A085KAD0 | Uncharacterized protein  |
| 203 | A0A085KAK4 | Peptidase M2 family protein (Peptidyl-dipeptidase)  | 203 | A0A3G2ULH4 | DUF1330 domain-containing protein                                    |
| 204 | A0A085JZY7 | ETC complex I subunit (ETC complex subunit I)   | 204 | A0A085K7Z5 | Membrane protein   |
| 205 | A0A3G2UKZ6 | Uncharacterized protein   | 205 | A0A3G2UMV2 | LPD7 domain-containing protein                                       |
| 206 | A0A3G2UKW0 | Type II toxin-antitoxin system Phd/YefM family antitoxin  | 206 | A0A3G2ULT0 | Uncharacterized protein  |
| 207 | A0A085KAV1 | Carboxylating nicotinate-nucleotide diphosphorylase   | 207 | A0A085K318 | Uncharacterized protein  |
| 208 | A0A291N0P8 | GLOBIN domain-containing protein  | 208 | A0A085K0K5 | Heavy metal resistance protein (Periplasmic heavy metal sensor)      |
| 209 | A0A085K7J1 | Signal recognition particle protein (Fifty-four homolog)  | 209 | A0A3G2UKY4 | DUF779 domain-containing protein                                     |
| 210 | A0A084ER82 | Peptide methionine sulfoxide reductase MsrA (Protein-methionine-S-oxide reductase)  | 210 | A0A084ERZ8 | Acyl carrier protein (Phosphopantetheine-binding protein)            |
| 211 | A0A085K319 | Asparaginase (L-asparaginase/GlutRNAGln amidotransferase subunit D)   | 211 | A0A085K344 | YciI family protein  |
| 212 | A0A085K908 | DUF3297 family protein (Glutathione peroxidase)   | 212 | A0A3G2UPY7 | ABC transporter permease   |
| 213 | A0A085K3G5 | Porphobilinogen deaminase (PBG)   | 213 | A0A085K2L9 | Membrane protein   |
| 214 | A0A3G2UPI8 | Glycerol kinase   | 214 | A0A3G2UPM5 | Aldehyde dehydrogenase   |
| 215 | A0A084EPR1 | Ribosomal silencing factor RsfS   | 215 | A0A3G2UV94 | MFS transporter  |
| 216 | A0A085K5J5 | Acetyl-CoA C-acyltransferase  | 216 | A0A3G2UZG5 | TetR family transcriptional regulator                                |
| 217 | A0A3G2UYN8 | Probable malate:quinone oxidoreductase  | 217 | A0A3G2ULN3 | RraA family protein  |
| 218 | A0A085K583 | tRNA uridine 5-carboxymethylaminomethyl modification enzyme MnmG (Glucose-inhibited division protein A)                                       | 218 | A0A3G2ULJ3 | 2-hydroxychromene-2-carboxylate isomerase                            |
| 219 | A0A085K6N4 | Fumarylacetoacetase   | 219 | A0A084EL78 | 17 kD surface antigen (Glycine zipper 2TM domain-containing protein) |
| 220 | A0A085KAL4 | Mannose-1-phosphate guanylyltransferase (Nucleotidyltransferase family protein)   | 220 | A0A084EHX2 | Uncharacterized protein  |
| 221 | A0A084ER14 | CTP synthase (EC 6.3.4.2) (Cytidine 5'-triphosphate synthase) (Cytidine triphosphate synthetase) (CTP synthetase) (CTPS) (UTP-ammonia ligase) | 221 | A0A3G2UX17 | Polysaccharide biosynthesis tyrosine autokinase                      |
| 222 | A0A085KAN0 | LuxR family transcriptional regulator (TatD family deoxyribonuclease)   | 222 | A0A3G2UQ13 | Uncharacterized protein  |
| 223 | A0A3G2UKB9 | Alpha/beta fold hydrolase   | 223 | A0A3G2UYK7 | Sulfate adenyltransferase subunit 2                                  |
| 224 | A0A085K5J6 | Nitronate monooxygenase   | 224 | A0A084ELP5 | Envelope stress response membrane protein PspB (Phage-shock protein) |
| 225 | A0A3G2UNZ2 | NAD(P)/FAD-dependent oxidoreductase   | 225 | A0A085K082 | Universal stress protein (Universal stress protein UspA)             |
| 226 | A0A3G2UUR1 | 3-methyl-2-oxobutanoate dehydrogenase (2-methylpropanoyl-transferring) subunit alpha  | 226 | A0A085JZ27 | Membrane protein (OmpA family protein)                               |
| 227 | A0A085K6N7 | Cobalamin biosynthesis protein CobW   | 227 | A0A085K2D3 | Threonine synthase   |
| 228 | A0A085KAG9 | Alanine racemase  | 228 | A0A3G2UNS7 | Conjugal transfer protein  |
| 229 | A0A3G2USH2 | Type I restriction endonuclease subunit S   | 229 | A0A085K2Q2 | Ribosomal RNA small subunit methyltransferase H                      |

(待续)

(续表 5)

|     |            |   |     |            |  |
|-----|------------|---|-----|------------|--|
| 230 | A0A085JYX9 | Glyoxalase (Lactoylglutathione lyase)   | 230 | A0A085K5D9 | Malic enzyme   |
| 231 | A0A3G2UQP5 | S-(hydroxymethyl)glutathione dehydrogenase  | 231 | A0A085K678 | Periplasmic serine endoprotease DegP-like  |
| 232 | A0A085K5J3 | 3-hydroxy-2-methylbutyryl-CoA dehydrogenase (SDR family NAD(P)-dependent oxidoreductase)  | 232 | A0A3G2UWP4 | Uncharacterized protein  |
| 233 | A0A084EKT4 | 50S ribosomal protein L25 (General stress protein CTC)                                    | 233 | A0A085K7F1 | Lipoprotein-releasing system ATP-binding protein LolD  |
| 234 | A0A085K9A4 | Enoyl-[acyl-carrier-protein] reductase [NADH]   | 234 | A0A3G2UYI6 | Biotin/lipoyl-binding protein  |
| 235 | A0A085K8Q7 | Lysine--tRNA ligase   | 235 | A0A085K5Z3 | GCN5 family acetyltransferase (GCN5-related N-acetyltransferase) (GNAT family N-acetyltransferase) (N-acetyltransferase) |
| 236 | A0A3G2UL15 | Molybdopterin dinucleotide-binding protein  | 236 | A0A085K6F9 | LysR family transcriptional regulator (Transcriptional regulator)  |
| 237 | A0A085K092 | M20/M25/M40 family metallo-hydrolase (Peptidase M20)                                      | 237 | A0A085K376 | 3-hydroxyacyl-[acyl-carrier-protein] dehydratase FabZ  |
| 238 | A0A085K9Q7 | Nucleoside triphosphate hydrolase   | 238 | A0A084EQU7 | Ubiquinol-cytochrome c reductase iron-sulfur subunit   |
| 239 | A0A085K1E3 | Glycosyl transferase family 1 (Glycosyltransferase family 1 protein)                      | 239 | A0A085K6Y6 | Acetyl-CoA C-acetyltransferase   |
| 240 | A0A084EJK6 | DUF2093 domain-containing protein   | 240 | A0A085JZ59 | Arylesterase (GDSL family lipase)  |
| 241 | A0A084ETG6 | MucR family transcriptional regulator (Transcriptional regulator)                         | 241 | A0A085K2Q1 | Transcriptional regulator MraZ   |
| 242 | A0A0J9D2S7 | Aspartyl/glutamyl-tRNA(Asn/Gln) amidotransferase subunit B (Asp/Glu-ADT subunit B)        | 242 | A0A084EQH0 | Glycine zipper 2TM domain-containing protein   |
| 243 | A0A085K3F8 | Ribose-phosphate pyrophosphokinase (RPPK)   | 243 | A0A3G2URY7 | FtsX-like permease family protein  |
| 244 | A0A085K3D2 | Histidinol-phosphate aminotransferase   | 244 | A0A085K292 | Autotransporter domain-containing protein (Serine protease)  |
| 245 | A0A291MVR2 | NADH-quinone oxidoreductase subunit F   | 245 | A0A3G2UNN3 | Sigma-70 family RNA polymerase sigma factor  |
| 246 | A0A3G2UWP0 | Acyl-CoA dehydrogenase  | 246 | A0A085K184 | DNA-binding response regulator (Transcriptional regulator) (Two-component system response regulator)                     |
| 247 | A0A084EQE2 | Elongation factor Ts (EF-Ts)  | 247 | A0A085K6B7 | Uncharacterized protein  |
| 248 | A0A0J9CVL9 | Cytoplasmic protein (DUF1013 domain-containing protein)                                   | 248 | A0A085K714 | Efflux transporter outer membrane subunit (RND transporter)  |
| 249 | A0A085K5Z2 | Membrane protein (OmpW family protein) (Outer membrane protein W)                         | 249 | A0A3G2UNU8 | YnbE family lipoprotein  |
| 250 | A0A3G2UYD7 | Ribosomal protein S12 methylthiotransferase RimO (S12 MTTase) (S12 methylthiotransferase) | 250 | A0A3G2ULB5 | Cbb3-type cytochrome c oxidase subunit   |
| 251 | A0A085KAF1 | GTPase Era  | 251 | A0A085JYY0 | SIMPL domain-containing protein  |
| 252 | A0A085K3C1 | Phosphoribosylformylglycinamide cyclo-ligase  | 252 | A0A3G2UNN5 | General stress protein   |
| 253 | A0A084ENT3 | Ribonucleoside-diphosphate reductase subunit beta   | 253 | A0A3G2UT87 | Tautomerase  |
| 254 | A0A085JZY1 | Glycine dehydrogenase (aminomethyl-transferring)  | -   | -          | -  |
| 255 | A0A3G2UP12 | Methylcrotonoyl-CoA carboxylase   | -   | -          | -  |
| 256 | A0A085K230 | 2-methylisocitrate lyase (Isocitrate lyase/ phosphoenolpyruvate mutase family protein)    | -   | -          | -  |
| 257 | A0A085KAS4 | 5-methyltetrahydrofolate-homocysteine methyltransferase                                   | -   | -          | -  |
| 258 | A0A085KAV2 | Peptidase S9 (S9 family peptidase)  | -   | -          | -  |

(待续)



(续表 5)

|     |            |  |   |   |   |
|-----|------------|--|---|---|---|
| 259 | A0A3G2UME0 | S9 family peptidase  | — | — | — |
| 260 | A0A085JZ23 | Ribonuclease   | — | — | — |
| 261 | A0A084EPY4 | Peptide deformylase (PDF)  | — | — | — |
| 262 | A0A085K2B6 | 50S ribosomal protein L5   | — | — | — |
| 263 | A0A085K1D3 | ATP synthase subunit delta (ATP synthase F(1) sector subunit delta) (F-type ATPase subunit delta) (F-ATPase subunit delta)               | — | — | — |
| 264 | A0A085K6E7 | Cysteine desulfurase   | — | — | — |
| 265 | A0A085K9G6 | 2,3-bisphosphoglycerate-dependent phosphoglycerate mutase (BPG-dependent PGAM) (PGAM) (Phosphoglyceromutase) (dPGM)                      | — | — | — |
| 266 | A0A085K5J2 | Acetyl-CoA C-acyltransferase   | — | — | — |
| 267 | A0A085K3B7 | Helicase   | — | — | — |
| 268 | A0A3G2UP86 | Amidohydrolase   | — | — | — |
| 269 | A0A3G2UM60 | CoA ester lyase  | — | — | — |
| 270 | A0A3G2UNi9 | DUF952 domain-containing protein   | — | — | — |
| 271 | A0A084ER03 | Acyl dehydratase (Dehydratase) (MaoC family dehydratase)   | — | — | — |
| 272 | A0A085KA38 | Transketolase  | — | — | — |
| 273 | A0A085K264 | Aspartate-tRNA(Asp/Asn) ligase (EC 6.1.1.23) (Aspartyl-tRNA synthetase) (AspRS) (Non-discriminating aspartyl-tRNA synthetase) (ND-AspRS) | — | — | — |
| 274 | A0A085K8P6 | MBL fold metallo-hydrolase   | — | — | — |
| 275 | A0A084ECG8 | 50S ribosomal protein L20  | — | — | — |
| 276 | A0A3G2V108 | Argininosuccinate lyase (ASAL)   | — | — | — |
| 277 | A0A085K0X5 | LexA repressor   | — | — | — |
| 278 | A0A084EH78 | ATP-binding protein (Putative ATPase)  | — | — | — |
| 279 | A0A084ES93 | 2-nitropropane dioxygenase (2-nitropropane dioxygenase-like enzyme)  | — | — | — |
| 280 | A0A085K044 | Electron transfer flavoprotein subunit alpha (Electron transfer flavoprotein subunit beta)   | — | — | — |
| 281 | A0A085K538 | Peptidase C69 (TldD/PmbA family protein)   | — | — | — |
| 282 | A0A084EPE7 | 50S ribosomal protein L14  | — | — | — |
| 283 | A0A085K7E7 | Ribose 5-phosphate isomerase (Ribose 5-phosphate isomerase B)  | — | — | — |
| 284 | A0A085K717 | 3-oxoacyl-ACP synthase (3-oxoacyl-[acyl-carrier-protein] reductase)  | — | — | — |
| 285 | A0A085K6F3 | Branched-chain-amino-acid aminotransferase   | — | — | — |
| 286 | A0A085K959 | Metalloprotease TldD (Protease TldD)   | — | — | — |
| 287 | A0A085KA19 | Alginate_exp domain-containing protein   | — | — | — |
| 288 | A0A0J9CY82 | Electron transfer flavoprotein subunit beta (Electron transfer flavoprotein subunit beta/FixA family protein)                            | — | — | — |
| 289 | A0A085K152 | Uncharacterized protein  | — | — | — |
| 290 | A0A085K2D8 | Ribonuclease PH (RNase PH)   | — | — | — |
| 291 | A0A084EPP4 | 50S ribosomal protein L17  | — | — | — |
| 292 | A0A085KAG3 | Methylmalonyl-CoA mutase   | — | — | — |
| 293 | A0A3G2UQG4 | Acyl-CoA dehydrogenase   | — | — | — |

(待续)

(续表 5)

|     |            |   |   |   |   |
|-----|------------|---|---|---|---|
| 294 | A0A3G2UTU5 | Sigma-54-dependent Fis family transcriptional regulator   | - | - | - |
| 295 | A0A084EHW5 | Aspartyl/glutamyl-tRNA(Asn/Gln) amidotransferase subunit C (Asp/Glu-ADT subunit C)  | - | - | - |
| 296 | A0A3G2UQS8 | N-acetyl-gamma-glutamyl-phosphate reductase (AGPR)  | - | - | - |
| 297 | A0A085K7R0 | Uracil-DNA glycosylase (UDG)  | - | - | - |
| 298 | A0A085K3G0 | Glutamate-tRNA ligase   | - | - | - |
| 299 | A0A084EQ83 | Ferredoxin  | - | - | - |
| 300 | A0A3G2UMX9 | GatB/YqeY domain-containing protein   | - | - | - |
| 301 | A0A085K7A7 | CocE/NonD family hydrolase (Glutaryl-7-ACA acylase)   | - | - | - |
| 302 | A0A084ETQ3 | GTP-binding protein TypA (Translational GTPase TypA)  | - | - | - |
| 303 | A0A085KA41 | Alpha-L-rhamnosidase  | - | - | - |
| 304 | A0A3G2UR06 | MBL fold hydrolase  | - | - | - |
| 305 | A0A085K554 | DUF4167 domain-containing protein   | - | - | - |
| 306 | A0A3G2UUK6 | UPF0246 protein EBF16_04420   | - | - | - |
| 307 | A0A3G2USF9 | Transcriptional regulator   | - | - | - |
| 308 | A0A085K1G3 | Valine-tRNA ligase  | - | - | - |
| 309 | A0A085K5B9 | Shikimate dehydrogenase (NADP(+)) (SDH)   | - | - | - |
| 310 | A0A085K9B6 | Cytokinin riboside 5'-monophosphate phosphoribohydrolase  | - | - | - |
| 311 | A0A085K3I0 | Bifunctional purine biosynthesis protein PurH [Includes: Phosphoribosylaminoimidazolecarboxamide formyltransferase  | - | - | - |
| 312 | A0A177JX07 | Carbamoyl-phosphate synthase large chain  | - | - | - |
| 313 | A0A085K3J0 | Single-stranded DNA-binding protein (SSB)   | - | - | - |
| 314 | A0A085K9E6 | UPF0178 protein EBF16_12115   | - | - | - |
| 315 | A0A085K8S9 | Uncharacterized protein   | - | - | - |
| 316 | A0A3G2V0V0 | TPR_REGION domain-containing protein  | - | - | - |
| 317 | A0A3G2UNL7 | Bifunctional protein PutA [Includes: Proline dehydrogenase  | - | - | - |
| 318 | A0A084EPP3 | DNA-directed RNA polymerase subunit alpha (RNAP subunit alpha)  | - | - | - |
| 319 | A0A085K7E6 | Serine hydroxymethyltransferase (SHMT) (Serine methylase)   | - | - | - |
| 320 | A0A085KAG1 | Acyl-CoA carboxylase subunit beta (Methylmalonyl-CoA carboxyltransferase)   | - | - | - |
| 321 | A0A3G2V1J1 | Shikimate 5-dehydrogenase   | - | - | - |
| 322 | A0A0J9CZF5 | Acyl-CoA thioesterase   | - | - | - |
| 323 | A0A085KA49 | Phosphoribosylformylglycinamide synthase subunit PurQ (FGAM synthase)   | - | - | - |
| 324 | A0A085K5F7 | Phosphoheptose isomerase  | - | - | - |
| 325 | A0A3G2V241 | S9 family peptidase   | - | - | - |
| 326 | A0A085K5D7 | Bifunctional uridylyltransferase/uridylyl-removing enzyme (UTase/UR) (Bifunctional [protein-PII] modification enzyme) (Bifunctional nitrogen sensor protein) [Includes: [Protein-PII] uridylyltransferase (PII uridylyltransferase) (UTase) | - | - | - |
| 327 | A0A085K7D5 | Agmatine deiminase family protein   | - | - | - |

Note: -, No data.

### 2.4.2 差异蛋白的生物信息学分析

分别对表达差异蛋白在分子功能、生物学过程以及 KEGG 信号通路层面进行了分析,如图 6 所示。在分子功能方面,除了催化和结合功能外,有近 10% 的表达差异蛋白具有转运的功能,这说明在菲的环境下,细菌需要表达更多的转运蛋白将菲、无机盐等物质转运至胞内。表达差异蛋白参与的生物学过程主要集中在代谢过程、细胞生长和生物调控等方面。KEGG 信号通路分析结果显示大量的蛋白参与了各类物质的代谢和合成。根据以上结果推测鞘脂菌 SJTF-8 需要表达更多可在多样化环境中发挥代谢功能的蛋白、转运蛋白以及生物调控蛋白等来适应菲的胁迫。

### 2.4.3 差异调控蛋白的比较

分别对 DIA 和 DDA 测试结果中 A 组发现的表达差异调控蛋白做了统计分析,发现利用 DIA 技术定量到 39 个调控蛋白(CV 值 $\leq 20\%$ ),数量是 DDA 中调控蛋白(CV 值 $\leq 20\%$ )的 3 倍以上,证明了 DIA 技术方法对低丰度差异蛋白检出的优势。如表 6 所示,我们还发现在这 39 个调控蛋白中,有 21 个调控蛋白在 DDA 所有定量结果中虽然被检测到,但

CV 值均大于 20%,进一步说明 DDA 对低丰度蛋白定量检测的重现性差,这也更加凸显出 DIA 的方法对低丰度调控蛋白检测的优势。本研究的菲胁迫下 SJTF-8 蛋白质组的定量结果为鞘脂菌对菲的降解途径调控机制的研究提供了新线索和方向。

## 3 讨论与结论

DIA 技术的出现为高通量、全面地定量蛋白质组学开辟了一个新的领域,已被广泛应用于高等生物样品如人类体液、组织、小鼠/大鼠和斑马鱼等动物模型的研究中<sup>[15,23-24]</sup>。Lin 等对血清蛋白的研究发现,与 DDA 技术相比,DIA 法检测到的多肽和蛋白质的数量增加一倍且具有较好的重现性<sup>[25]</sup>;Muntel 等对 87 份尿液样品的检测中也发现,相较于 DDA 法,DIA 明显提高了检测通量以及定量能力,使得高效地发现和验证生物标记物成为可能<sup>[14]</sup>。但是在低等生物中 DIA 技术的应用研究较少,目前仅在幽门螺旋杆菌、结核分枝杆菌和酵母的研究中得到应用<sup>[26-28]</sup>。Selevsek 等将 DIA 运用于酵母的研究中,发现 DIA 技术不仅与多重反应监测技术有相当的精度、重复型和准确性,还允许在多个样品中定

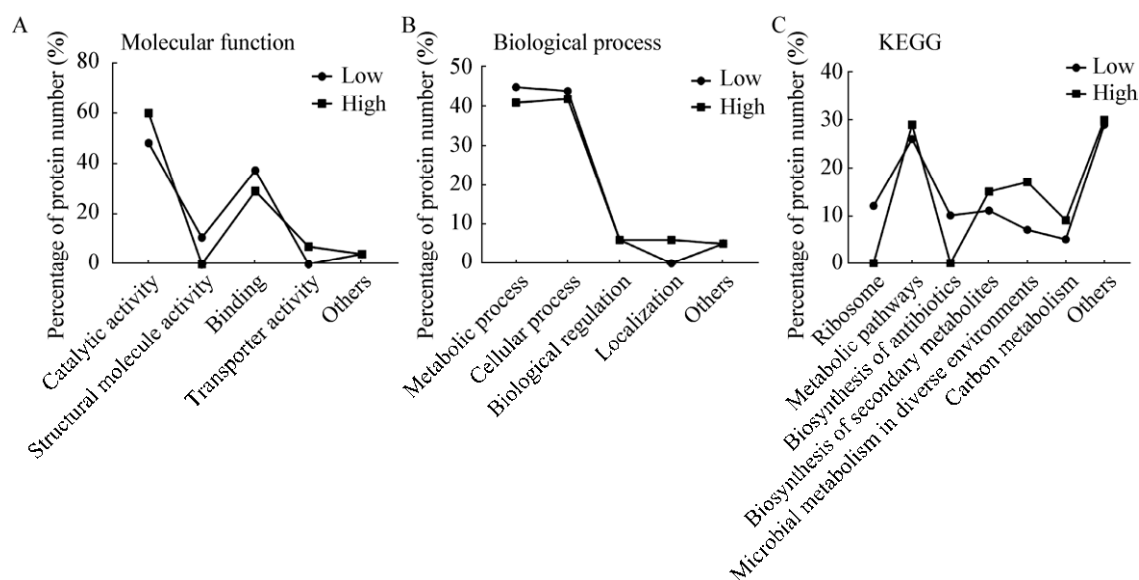


图 6 DDA 和 DIA 的共同得到的差异蛋白生物信息学分析

Figure 6 Bioinformatics analysis of differentially expressed proteins both from DDA and DIA

表 6 DDA 中 CV 值≥20%的 21 个调控蛋白  
Table 6 21 regulatory proteins with CV≥20% in DDA

| No. | Accessions | Protein names  |
|-----|------------|--|
| 1   | A0A3G2UX28 | Transcriptional regulator  |
| 2   | A0A3G2UTF0 | Type II toxin-antitoxin system ParD family antitoxin   |
| 3   | A0A3G2V104 | LysR family transcriptional regulator  |
| 4   | A0A3G2UPX2 | Transcriptional regulator  |
| 5   | A0A3G2UTK0 | MerR family transcriptional regulator  |
| 6   | A0A3G2UTM0 | PAS domain-containing sensor histidine kinase  |
| 7   | A0A085K5Z7 | Crp/Fnr family transcriptional regulator (cAMP-binding protein-catabolite transcription activator) |
| 8   | A0A085K957 | Nitrogen regulatory protein P-II 1 (P-II family nitrogen regulator)                                |
| 9   | A0A085K7C3 | Histidine kinase (Putative regulator of cell autolysis) (Sensor histidine kinase)                  |
| 10  | A0A085K417 | FadR family transcriptional regulator (GntR family transcriptional regulator)                      |
| 11  | A0A084EUT7 | Histidine phosphotransferase (Hpt domain-containing protein)                                       |
| 12  | A0A085K446 | LacI family DNA-binding transcriptional regulator (LacI family transcriptional regulator)          |
| 13  | A0A085K6A6 | FadR family transcriptional regulator (GntR family transcriptional regulator)                      |
| 14  | A0A085K0Y0 | Histidine kinase (Sensor histidine kinase)   |
| 15  | A0A085K5M0 | MarR family transcriptional regulator  |
| 16  | A0A085K4Q1 | ArsR family transcriptional regulator  |
| 17  | A0A085K9N9 | Uncharacterized protein  |
| 18  | A0A085K560 | DeoR family transcriptional regulator (PLP-dependent aminotransferase family protein)              |
| 19  | A0A085JZV3 | Chemotaxis protein CheY (DNA-binding response regulator) (Two-component system response regulator) |
| 20  | A0A3G2ULP2 | Glutaredoxin family protein  |
| 21  | A0A3G2UQ46 | Transcriptional regulator  |

量大量蛋白质，提高了传统方法的效率<sup>[27]</sup>。除此之外，Schubert 等对结核分枝杆菌的研究也让我们看到了将 DIA 运用到绝对定量蛋白质组学中的可能性<sup>[28]</sup>。

本研究选取低等生物鞘脂菌 SJTF-8 为分析对象，从缺失值、重现性和定性定量蛋白数目等方面比较了 DDA 和 DIA 技术的定量特点，发现 DIA 技术重现性和对低丰度蛋白的定量能力明显优于 DDA 技术。因此 DIA 在实际可用的表达差异蛋白检出方面具备明显优势，尤其有利于发现非胁迫下细胞诱导表达的低丰度调控蛋白。然而，DIA 相较于 DDA 技术在蛋白质组定性定量总体数量方面不具备显著优势，这可能是由于本实验以微生物蛋白质组为研究对象，鞘氨醇类菌的蛋白家族数量有限，远少于高等生物，现有的仪器在 DDA 模式下就可以采集到较为全面的蛋白信息。因此，对于蛋

白成分简单的样品，使用 DDA 采集模式不仅可以获得相当体量的蛋白质组定性定量结果，而且具有样品前处理及质谱采集方法设置简单等优势。科研人员可以根据研究对象的特点和研究目的合理选择 DDA 和 DIA 技术进行蛋白质组相对定量分析。

鞘氨醇类菌(包含鞘氨醇单胞菌、鞘脂菌、鞘氨醇盒菌以及新鞘脂菌属)是最常见的非降解菌之一，在微生物修复多环芳烃的污染方面具有重要意义<sup>[29]</sup>。人们对于鞘氨醇类菌降解菲的代谢途径已做了较为广泛的研究<sup>[3]</sup>。但是，关于多环芳烃降解的调控机制、代谢产物聚集等方面的机理尚不清楚<sup>[1]</sup>。鞘氨醇类菌的蛋白质组研究，可以为其在微生物修复方面的基因调控机制提供清晰的思路。然而，关于鞘氨醇类菌在多环芳烃环境下的蛋白质组学研究则少之又少，而且鉴定到的蛋白数量也有一定限制<sup>[1,30-31]</sup>。

本实验以鞘氨醇类菌中鞘脂菌为研究对象,通过 DDA 和 DIA 两种技术方法进行非胁迫下鞘脂菌蛋白质组非标记相对定量分析,共得到 580 个表达差异蛋白,更完整地优化出鞘脂菌 *Sphingobium yanoikuyae* SJTF-8 在非胁迫下表达差异的蛋白。另外,GO 和 KEGG 分析表明表达差异蛋白在细胞代谢、转运和调控等方面都发挥一定功能,为深入研究鞘脂菌的多环芳烃降解过程提供了理论依据,也为更好地治理有机物污染提供了新的研究思路。

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## 稿件书写规范

### 专论与综述论文的撰写要点

专论与综述是本刊重要栏目之一, 主要反映国内外微生物学及相关领域学科研究最新成果和进展, 其内容要求新颖丰富, 观点明确, 论述恰当, 应包含作者自己的工作内容和见解。因此, 作者在动笔之前必须明确选题, 一般原则上应选择在理论和实践中具有重要意义的学科专题进行论述。围绕专题所涉及的各个方面, 在综合分析和评价已有资料基础上提出其演变规律和趋势, 即掌握其内在的精髓, 深入到专题研究的本质, 论述其发展前景。作者通过回顾、观察和展望, 提出合乎逻辑并具有启迪性的看法和建议。另外, 作者也可以采用以汇集文献资料为主的写作方法, 辅以注释, 客观而有少量评述, 使读者对该专题的过去、现在和将来有一个全面、足够的认识。

需要特别说明的是: (1) 我刊要求作者投稿时在正文前写上主要作者专业和研究背景的简介, 并指出自己的工作(已发表的文章)在综述中的体现, 同时请在稿件中用不同颜色标出来。(2) 在专论与综述中引用的文献应该主要是近 5 年国内外正式发表的研究论文, 引用文献数量不限。