

简报

## 酵母基因克隆受体菌的构建

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近年来,一些国家的实验室采用酵母菌重组DNA技术将外源淀粉酶基因引入酿酒酵母中,构建成具有分解淀粉能力的工程菌株,其中酵母糖化酶基因是重要的来源之一<sup>[1]</sup>。大部分酿酒酵母携带糖化酶基因的抑制基因(*INH1*)<sup>[2]</sup>,为了在酿酒酵母中克隆和表达糖化酵母糖化酶基因(*STA*),需要构建适宜的受体菌,其遗传标记应为/*leu2* *sta<sup>+</sup>**inh<sup>+</sup>*。

本文报道了糖化酶基因克隆受体菌的构建,同时还得到一批带有不同选择标记的糖化酵母杂种菌株。

## 实验与结果

已知*INH1*与*STA*基因不连锁,可通过酵母菌杂交技术,构建适宜的受体菌<sup>[3]</sup>。为了尽快得到所需的受体菌(*trp1 sta<sup>+</sup>inh<sup>+</sup>*),我们设计了两步杂交路线(见图1)。

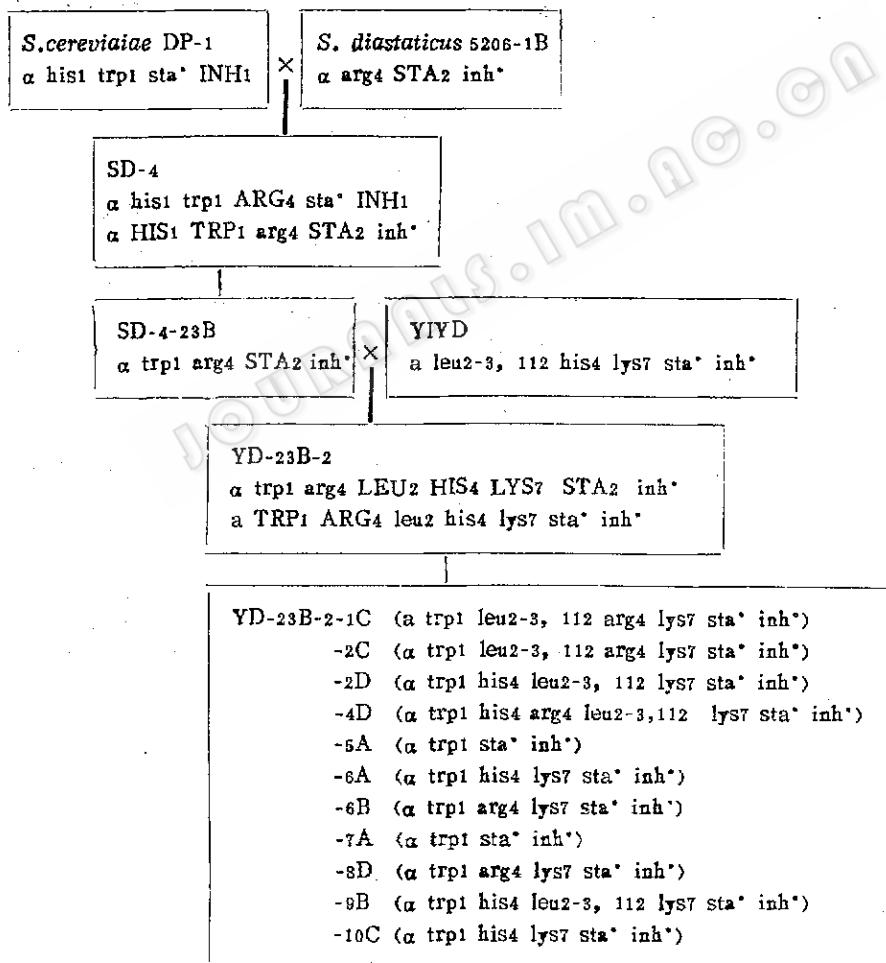


图1 糖化酶基因克隆受体菌株构建的杂交谱系

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首先用酿酒酵母DP1(P.P.Slonimski赠)与糖化酵母5206-1B(杭州大学李桃生转赠)进行杂交,经28℃培养后,用显微操作器从杂交群体中挑出6个杂合子。其中SD-4<sup>-</sup>经纯培养后再在生孢培养基(1%KAC、1%葡萄糖、0.25%酵母粉及2%琼脂粉)上28℃培养1天后,室温放置24h,用显微操作器解剖4孢子子囊10个,经四分体分析<sup>[4]</sup>得到一批带有trp<sup>1</sup>及其它缺陷标记的糖化酵母杂种后代(见表1)。

从上述得到的糖化酵母杂种后代中选出SD-4-23B( $\alpha$  trp<sup>1</sup> arg<sup>4</sup> STA<sub>2</sub> inh<sup>+</sup>)菌株再和糖化酵母YIYD( $\alpha$  leu<sup>2</sup> his<sup>4</sup> lys<sup>7</sup> sta<sup>+</sup> inh<sup>+</sup>)(Ichiro Yamashita赠)杂交,用显微操作器挑出3个杂合子,其中之-YD-23B-2杂合子在生孢子培养基上培养后,用显微操作器解剖四孢子子囊,得到了一批杂种后代(见表2),经测定这些后代的营养要求和淀粉水解能力,得到了所需的受体菌。其中YD-23B-2-1C、2C、2D、4D、5A、6A、6B、7A、8D、9B和10C这11株菌均带有trp<sup>1</sup>缺陷标记和sta<sup>+</sup> inh<sup>+</sup>基因型,适宜于带

表1 DP1×5206-1B杂种后代的遗传特性

Strain	Genotype
SD-4-12B	MAT $\alpha$ his STA <sub>2</sub> inh <sup>+</sup>
-12C	MAT $\alpha$ arg <sup>4</sup> STA <sub>2</sub> inh <sup>+</sup>
-13B	MAT $\alpha$ trp <sup>1</sup> arg <sup>4</sup> STA <sub>2</sub> inh <sup>+</sup>
-13C	MAT $\alpha$ his <sup>1</sup> STA <sub>2</sub> inh <sup>+</sup>
-14B	MAT $\alpha$ trp <sup>1</sup> arg <sup>4</sup> STA <sub>2</sub> inh <sup>+</sup>
-14C	MAT $\alpha$ his <sup>1</sup> STA <sub>2</sub> inh <sup>+</sup>
-16A	MAT $\alpha$ his <sup>1</sup> STA <sub>2</sub> inh <sup>+</sup>
-16B	MAT $\alpha$ trp <sup>1</sup> his <sup>1</sup>
-16C	MAT $\alpha$ arg <sup>4</sup>
-16D	MAT $\alpha$ trp <sup>1</sup> arg <sup>4</sup>
-18C	MAT $\alpha$ arg <sup>4</sup> STA <sub>2</sub> inh <sup>+</sup>
-19A	MAT $\alpha$ his <sup>1</sup>
-19C	MAT $\alpha$ arg <sup>4</sup>
-19D	MAT $\alpha$ trp <sup>1</sup> his <sup>1</sup>
-22B	MAT $\alpha$ his <sup>1</sup> STA <sub>2</sub> inh <sup>+</sup>
-22C	MAT $\alpha$ trp <sup>1</sup> his <sup>1</sup> STA <sub>2</sub> inh <sup>+</sup>
-23A	MAT $\alpha$ trp <sup>1</sup> arg <sup>4</sup>
-23B	MAT $\alpha$ trp <sup>1</sup> arg <sup>4</sup> STA <sub>2</sub> inh <sup>+</sup>
-23C	MAT $\alpha$ his <sup>1</sup>
-23D	MAT $\alpha$ his <sup>1</sup>
-24D	MAT $\alpha$ trp <sup>1</sup> his <sup>1</sup> STA <sub>2</sub> inh <sup>+</sup>
-28A	MAT $\alpha$ his <sup>1</sup> STA <sub>2</sub> inh <sup>+</sup>
-28D	MAT $\alpha$ trp <sup>1</sup> arg <sup>4</sup> STA <sub>2</sub> inh <sup>+</sup>

表2 SD-4-23B×YIYD杂种后代的遗传特性

Strain	Genotype
YD-23B-2-1A	MAT $\alpha$ lys <sup>7</sup> sta <sup>+</sup> inh <sup>+</sup>
-1B	MAT $\alpha$ trp <sup>1</sup> his <sup>4</sup> leu <sup>2-3</sup> , 112 STA <sub>2</sub> inh <sup>+</sup>
-1C	MAT $\alpha$ trp <sup>1</sup> leu <sup>2-3</sup> , 112 arg <sup>4</sup> lys <sup>7</sup> sta <sup>+</sup> inh <sup>+</sup>
-1D	MAT $\alpha$ his <sup>4</sup> arg <sup>4</sup> lys <sup>7</sup> STA <sub>2</sub> inh <sup>+</sup>
-2A	MAT $\alpha$ STA <sub>2</sub> inh <sup>+</sup>
-2B	MAT $\alpha$ arg <sup>4</sup> STA <sub>2</sub> inh <sup>+</sup>
-2C	MAT $\alpha$ trp <sup>1</sup> leu <sup>2-3</sup> , 112 arg <sup>4</sup> lys <sup>7</sup> sta <sup>+</sup> inh <sup>+</sup>
-2D	MAT $\alpha$ trp <sup>1</sup> his <sup>4</sup> leu <sup>2-3</sup> , 112 lys <sup>7</sup> sta <sup>+</sup> inh <sup>+</sup>
-4A	MAT $\alpha$ lys <sup>7</sup> STA <sub>2</sub> inh <sup>+</sup>
-4B	MAT $\alpha$ sta <sup>+</sup> inh <sup>+</sup>
-4C	MAT $\alpha$ trp <sup>1</sup> his <sup>4</sup> leu <sup>2-3</sup> , 112 arg <sup>4</sup> STA <sub>2</sub> inh <sup>+</sup>
-4D	MAT $\alpha$ trp <sup>1</sup> his <sup>4</sup> leu <sup>2-3</sup> , 112 arg <sup>4</sup> lys <sup>7</sup> sta <sup>+</sup> inh <sup>+</sup>
-5A	MAT $\alpha$ trp <sup>1</sup> sta <sup>+</sup> inh <sup>+</sup>
-5B	MAT $\alpha$ his <sup>4</sup> leu <sup>2-3</sup> , 112 arg <sup>4</sup> STA <sub>2</sub> inh <sup>+</sup>
-5C	MAT $\alpha$ his <sup>4</sup> leu <sup>2-3</sup> , 112 lys <sup>7</sup> sta <sup>+</sup> inh <sup>+</sup>
-5D	MAT $\alpha$ trp <sup>1</sup> arg <sup>4</sup> lys <sup>7</sup> STA <sub>2</sub> inh <sup>+</sup>
-6A	MAT $\alpha$ trp <sup>1</sup> his <sup>4</sup> lys <sup>7</sup> sta <sup>+</sup> inh <sup>+</sup>
-6B	MAT $\alpha$ trp <sup>1</sup> arg <sup>4</sup> lys <sup>7</sup> sta <sup>+</sup> inh <sup>+</sup>
-6D	MAT $\alpha$ his <sup>4</sup> leu <sup>2-3</sup> , 112 arg <sup>4</sup> STA <sub>2</sub> inh <sup>+</sup>
-7A	MAT $\alpha$ trp <sup>1</sup> sta <sup>+</sup> inh <sup>+</sup>
-7C	MAT $\alpha$ his <sup>4</sup> leu <sup>2-3</sup> , 112 arg <sup>4</sup> sta <sup>+</sup> inh <sup>+</sup>
-7D	MAT $\alpha$ lys <sup>7</sup> STA <sub>2</sub> inh <sup>+</sup>
-8A	MAT $\alpha$ his <sup>4</sup> leu <sup>2-3</sup> , 112 STA <sub>2</sub> inh <sup>+</sup>
-8D	MAT $\alpha$ trp <sup>1</sup> arg <sup>4</sup> lys <sup>7</sup> sta <sup>+</sup> inh <sup>+</sup>
-9A	MAT $\alpha$ leu <sup>2-3</sup> , 112 STA <sub>2</sub> inh <sup>+</sup>

接表 2

Strain	Genotype
-9B	MAT <sub>a</sub> trp1 his4 leu2-3, 112 lys7 sta° inh°
-10A	MAT <sub>a</sub> his4 leu2-3, 112 arg4 sta° inh°
-10B	MAT <sub>a</sub> trp1 arg4 STA2 inh°
-10C	MAT <sub>a</sub> trp1 his4 lys7 sta° inh°
-10D	MAT <sub>a</sub> leu2-3, 112 lys7 STA2 inh°
-8A	MAT <sub>a</sub> trp1 STA2 inh°
-8C	MAT <sub>a</sub> his4 leu2-3, 112 lys7 sta° inh°
-8D	MAT <sub>a</sub> his4 leu2-3, 112 arg4 lys7 sta° inh°

TRP1 选择标记的酵母穿梭质粒的转化；此外 YD-23B-2-1C、2C、2D、3C、3D、4D、5C、7C、9B 及 10A 这 10 株菌带有 leu2 Sta° inh° 的基因型，可用于带 LEU2 选择标记的酵母穿梭质粒的转化。以上两类菌株均可用以酵母糖化酶基因克隆的受体菌。

在表 2 中还有一些带 trp1 STA2 和 leu2 STA2 基因型的菌株，它们可以分泌胞外糖化酶，具有淀粉发酵能力，还适宜于 YE<sub>p</sub> 型和 YR<sub>p</sub> 型质粒的转化。这是一类新的酵母受体菌，通过引入外源 α- 淀粉酶基因，可望构建成分解淀粉酿酒酵母。

我们从两次杂交的子囊后代中得到一批带有不同缺陷标记的糖化酵母杂种后代，用淀粉水解圈半径测量，Durbam 氏法和二氧化碳减轻法<sup>[5]</sup> 测定了它们的淀粉发酵速率和淀粉发酵力（结果未给出），结果表明这些菌株的淀粉发酵速率差异较大，其中 YD-23B-2-4A、9A、10D 最快，

且淀粉发酵力也比其他菌株高，但和葡萄糖发酵能力相比还远不如，这和其它的报道结果相近。

## 讨 论

为了得到带 trp1 缺陷标记的酵母糖化酶基因克隆受体菌，我们采用了图 1 的二步杂交法，它有以下优点：路线简洁、方法简便、工作量小、得到的受体菌种类较多，在第二步杂交后，那些带 trp1 缺陷标记且无淀粉发酵力的菌株即为所需菌株。

在 DP1 × 5206-1B 杂交中，亲本 5206-1B 带有 POF（编码阿魏酸脱羧酶）基因，DP1 中此基因为隐性（pof），已知 STA 与 POF 基因不连锁，此杂交得到的杂种后代中肯定会出现 STA POF 基因型的杂种菌株，它能发酵糊精而不产生酚醛气味，因此适合于低卡值啤酒的发酵。因此这为我们进一步筛选和培育能酿造低卡啤酒的菌种打下了基础。

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## Construction of Recipient Strains for Cloning of Yeast Glucose Amylase Gene

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The recipient strains for cloning of yeast glucose amylase gene have been constructed by means of micromanipulation technique. One of the segregants with auxotrophic marker (*trp1*), SD-4-23b (a *trp1 arg4 STA2 inh<sup>o</sup>*), was obtained from the cross between *S.cerevisiae* DP-1 (*his1 trp1 sta<sup>o</sup> INH1*) and *S.diastaticus* 5206-1B (a *arg4 STA2 inh<sup>o</sup>*). Then the segregant was hybridized with *S.cerevisiae* YIYD (a *leu2-3, 112 his4 lys7 sta<sup>o</sup> inh<sup>o</sup>*). Eleven recipient strains (*trp1 sta<sup>o</sup> inh<sup>o</sup>*) derived from the latter cross are suitable for the transformation of *E.coli*-yeast shuttle plasmid pCN60 (TRP1) and cloning of glucose amylase gene. Some of the segregants with different auxotrophic markers and STA2 gene which were capable of fermenting starch more rapidly were also obtained.

### Key words

Recipient strains; cross; auxotrophic marker; *S.diastaticus*